Drug and Vaccine Development for the Treatment and Prevention of Urinary Tract Infections

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ABSTRACT Urinary tract infections (UTI) are among the most common bacterial infections in humans, affecting millions of people every year. UTI cause significant morbidity in women throughout their lifespan, in infant boys, in older men, in individuals with underlying urinary tract abnormalities, and in those that require long-term urethral catheterization, such as patients with spinal cord injuries or incapacitated individuals living in nursing homes. Serious sequelae include frequent recurrences, pyelonephritis with sepsis, renal damage in young children, pre-term birth, and complications of frequent antimicrobial use including high-level antibiotic resistance and Clostridium difficile colitis. Uropathogenic E. coli (UPEC) cause the vast majority of UTI, but less common pathogens such as Enterococcus faecalis and other enterococci frequently take advantage of an abnormal or catheterized urinary tract to cause opportunistic infections. While antibiotic therapy has historically been very successful in controlling UTI, the high rate of recurrence remains a major problem, and many individuals suffer from chronically recurring UTI, requiring long-term prophylactic antibiotic regimens to prevent recurrent UTI. Furthermore, the global emergence of multi-drug resistant UPEC in the past ten years spotlights the need for alternative therapeutic and preventative strategies to combat UTI, including anti-infective drug therapies and vaccines. In this chapter, we review recent advances in the field of UTI pathogenesis, with an emphasis on the identification of promising drug and vaccine targets. We then discuss the development of new UTI drugs and vaccines, highlighting the challenges these approaches face and the need for a greater understanding of urinary tract mucosal immunity.

THE URGENT NEED FOR NEW THERAPIES AND VACCINES

Urinary tract infections (UTI) are one of the most common bacterial infections, with roughly eleven-million cases reported in the U.S. each year that cost an estimated $5 billion annually (1, 2). More than one in every two women will experience at least one UTI in her lifetime, and nearly one in three women will have received antibiotic treatment for a UTI before age 24 (3, 4). The clinical manifestations of symptomatic UTI include infection-induced inflammation of the urethra (urethritis), urinary bladder (cystitis), and kidneys (pyelonephritis) and are diagnosed by the presence of high levels of bacteria in the urine (bacteriuria) with concomitant symptoms. Symptoms of cystitis include frequent urination, burning sensation and pain during urination.

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(dysuria), suprapubic pain and/or lower abdominal discomfort, and cloudy and/or bloody, foul-smelling urine. Symptoms of pyelonephritis include the presence of bacteriuria and pyuria (white blood cells in the urine) that is accompanied by flank pain and fever, but may or may not include other symptoms of cystitis. The vast majority of UTI manifest as cystitis and urethritis, affecting primarily the lower urinary tract, but this can potentially lead to bacterial ascension to the kidneys and pyelonephritis, particularly in pregnant women, diabetics, and children with vesicoureteral reflux (VUR) \(^\text{(5, 6)}\). As a result, renal scarring and loss of function is a potentially serious complication of any UTI, particularly in infants, where diagnosis of UTI may be delayed.

UTI are not only common, but also highly recurrent. In particular, sexually active women, the elderly, and prepubertal children are highly susceptible to chronically recurrent UTI, resulting in increased use of antibiotics and negatively affecting quality of life \(^\text{(3)}\). Approximately 20% to 30% of adult women with an initial UTI will experience a recurrence within 3–4 months \(^\text{(7)}\). In children, about one in three experiencing a UTI before the age of one will experience a recurrence within three years, and 18% will have a recurrence within a few months \(^\text{(8)}\). Uncomplicated UTI, which are infections that are not associated with urethral instrumentation or abnormal anatomy or physiology of the urinary tract, predominantly affect women, young children, and the elderly. Risk factors for uncomplicated cystitis in adult women include environmental factors such as frequent sexual activity, exposure to spermicides, menopause, and a history of childhood UTI, as well as genetic factors such as Toll-like receptor polymorphisms and a maternal history of UTI \(^\text{(9, 10)}\). In contrast, patients at risk for what is termed “complicated” UTI include patients with spinal cord injuries, patients undergoing urethral catheterization, diabetics, and individuals with underlying urologic abnormalities such as vesicoureteral reflux (VUR) \(^\text{(3)}\). Uropathogenic *Escherichia coli* (UPEC) cause 85% or more of uncomplicated UTI cases, while other Gram-negative rods and Gram-positive cocci, such as *Staphylococcus saprophyticus* and enterococci, are responsible for the remaining 5% to 15% of cases \(^\text{(11)}\).

The epidemiology of UTI changes significantly in the health care environment. Urethral catheterization is strongly associated with UTI, and the risk of infection increases with the length of catheterization \(^\text{(12)}\). Catheter-associated UTI (CAUTI) account for 30% to 40% of health care-associated infections in the United States, making them the most common nosocomial infection, with more than one-million cases occurring yearly in hospitals and nursing homes \(^\text{(13)}\). Although enterococci contribute only minimally to the burden of uncomplicated UTI, data from a national surveillance network of 463 hospitals in the United States revealed that 15% of CAUTI are caused by enterococci, second among bacterial isolates only to *E. coli* (21%) \(^\text{(14)}\). Furthermore, CAUTI affect both sexes, as long-term urinary catheterization of both men and women almost invariably leads to detection of bacteria in the urine (bacteriuria) and carries a daily risk of 3% to 7% for the development of symptomatic CAUTI \(^\text{(15)}\). While CAUTI is most often asymptomatic, the high incidence in catheterized patients greatly increases their risk for relatively rare but serious sequelae such as bacteremia, urosepsis, and death \(^\text{(16)}\). Furthermore, CAUTI serve as reservoirs for the dissemination of antimicrobial-resistant nosocomial pathogens in the health care environment \(^\text{(17)}\).

Although antibiotic therapy has historically been very successful in combating both uncomplicated and complicated UTI, many individuals suffer from chronically recurrent cystitis, requiring long-term antibiotic prophylaxis \(^\text{(18)}\). Furthermore, the widespread use of antibiotics has led to accelerating antibiotic resistance and the emergence and spread of multidrug-resistant (MDR) uropathogens \(^\text{(19)}\). As early as 1957, Weyrauch and colleagues foresaw this problem in their discussion of the results of a UTI vaccine trial \(^\text{(20)}\). In their study, they found that intramuscular injection with heat-killed *E. coli* was protective or partially protective against pyelonephritis in 12 of 16 rabbits. However, unvaccinated rabbits treated with prophylactic tetracycline were completely resistant to pyelonephritis, leading the authors to predict that prophylactic antibiotic treatment would remain the best strategy for preventing UTI in humans. Despite the authors’ admonition that “every effort must be made to avoid indiscriminate use” of antibiotic prophylaxis in order to prevent resistance, drug-resistant UTI has exploded into a major public health concern. For instance, a recent five-year nested case-control study of drug resistance in uncomplicated febrile UTI in adults found that 12% of patients with UPEC UTI had fluoroquinolone-resistant urine cultures; fluoroquinolone use in the previous six months was a significant independent risk factor for being afflicted with a fluoroquinolone-resistant UTI \(^\text{(21)}\). In another study, more than 9,000 patient urine samples were analyzed for drug resistance; of the samples containing uropathogens, 22.1% were multi-drug resistant (resistant to third-generation cephalosporins, ciprofloxacin, and aminoglycosides) \(^\text{(22)}\). In the past decade, the *E. coli* clone O25:H4-ST131 (*E. coli* ST131) emerged globally

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as an important MDR UPEC strain (23). Unlike other antibiotic-resistant UPEC strains, ST131 is highly virulent in the urinary tract and is not only found in health care settings, but is also isolated from the community (19, 24).

In summary, urinary tract infection is a significant cause of morbidity in women throughout their lifespan, in infant boys, and in older men. Serious sequelae include frequent recurrences, pyelonephritis with sepsis, renal damage in young children, pre-term birth, and complications of frequent antimicrobial use including high-level antibiotic resistance and Clostridium difficile colitis (25, 26). High recurrence rates and increasing antimicrobial resistance among uropathogens threaten to greatly increase the economic burden of this common infection. It has become increasingly evident that prophylactic use of antibiotics to prevent UTI is not a sustainable solution. The high incidence and recurrence rate of UTI, along with the rapid rise of MDR uropathogens and CAUTI, necessitate new drugs and vaccine therapies for the prevention of these infections. In this chapter, we will review UTI pathogenesis, focusing on UPEC as a model organism for uncomplicated UTI and Enterococcus faecalis as a model organism for complicated, CAUTI. We will then describe the development of anti-virulence therapies, including new classes of small-molecule inhibitors that target uropathogenic virulence factors. Finally, we will discuss vaccines for the prevention of recurrent UTI, including both whole cell and specific-antigen vaccines. It is our hope that this chapter will draw attention to recent advances in the field of UTI therapeutics while highlighting specific topics that require further study.

RECENT DISCOVERIES IN UTI PATHOGENESIS

Since 90% of symptomatic UTI present as simple cystitis/urethritis, an ideal drug or vaccine target would be one that is critical for both establishing and maintaining bladder colonization, thus preventing UTI altogether. Quantum-leap advances in molecular-biological and imaging technologies in the past 15 years, along with the maturation of genomic science, have led to an unprecedented expansion of our understanding of UTI pathogenesis, and to the identification of previously unknown virulence mechanisms. For example, we now know that UPEC invade bladder epithelial cells and have the capacity to rapidly replicate within the cytoplasm of superficial facet cells of the bladder urothelium, producing between 10,000 and 100,000 daughter cells from a single invasive bacterium within 12–16 hours (27–30). The establishment of this protected intracellular niche, known as the intracellular-bacterial community or IBC, helps UPEC gain a foothold in the lower urinary tract. Although discovered in mice, exfoliated bladder epithelial cells containing IBCs have been observed in urine sediments obtained from women and children with recurrent UTI, but not in healthy controls or in cases of UTI caused by Gram-positive pathogens (31–33), indicating that the murine model is a relevant and powerful tool for studying UTI pathogenesis. Indeed, mice are naturally susceptible to UPEC UTI and recapitulate many of the known characteristics of UTI in humans (34, 35).

While the translation of findings from animal models to the clinic is always fraught with difficulties, a major challenge in developing new therapeutics is not just the species differences between mice and humans in our animal models, but also the fact that these experimental infections are typically performed in naive animals. Experimental models of UTI in naive mice and primates have revealed similar pathogenic mechanisms of both cystitis and pyelonephritis (36–41). In contrast, recently developed models of recurrent UTI and post-menopausal UTI in mice have found urogenital mucosal immune responses very different from what is seen in naive mice (42–45). The epidemiology of UTI suggests that in a damaged or sensitized mucosal environment the requirements for bacterial virulence factors are diminished, potentially making any therapeutic intervention that targets the bacteria a tremendous challenge. In this section, we will briefly summarize our current knowledge of acute UTI pathogenesis in the latest animal models of uncomplicated UTI and CAUTI, highlighting the bacterial factors and host-pathogen processes that are promising drug and vaccine targets (Fig. 1 and Table 1).

Uncomplicated UTI: UPEC

UPEC adhesins

Whole-genome sequencing of several “prototypical” UPEC strains in the past 10 years has revealed the presence in each strain of multiple known and putative adhesins, a number of which have been demonstrated to contribute to UPEC’s ability to colonize the urinary tract. These include adhesive fibers called pili (fimbriae). A molecular machine known as the chaperone-usher pathway (CUP) mediates the assembly of pili on the bacterial outer membrane of diverse genera of Gram-negative bacteria (46–51) (detailed in “Structure, Function, and Assembly of Adhesive Organelles by
Uropathogenic Bacteria” by Thanassi et al.) and summarized in Fig. 2). Pili are long fibers that extend beyond the bacterial capsule. They contain adhesins at their tips that are thought to play an important role in host-pathogen interactions (52). Each sequenced UPEC strain encodes a multitude of CUP operons (53–55). For example, the cystitis strain UTI89 encodes 10 CUP operons, but of those that are broadly conserved among UPEC isolates, only two, type 1 and P pili, have so far been strongly implicated in UTI pathogenesis (Fig. 2). CUP adhesins are known to recognize specific receptors with stereochemical specificity. For example, FimH, the tip adhesin of the type 1 pilus, has been shown to bind mannosylated glycoproteins (56–58), as well as N-linked oligosaccharides on α3 and β1 integrins (59), and the pattern-recognition receptor Toll-like receptor 4 (TLR4) (60), all of which are expressed on the luminal surface of human and murine bladder epithelium. In contrast, the P pilus adhesin, PapG, is known to bind to Gal-α-1,4-Gal in globosides in the human kidney (61).

Type 1 pili

Several lines of evidence point to the type 1 pilus as a critical virulence factor in the establishment of UTI by UPEC in humans. Type 1 pili have been shown to be expressed during human UTI. A number of studies investigating whether type 1 pili are expressed by UPEC during UTI have found that the frequency of positive type 1 pilus immunostaining in urine sediments from women with acute UTI ranged from 40% to 76% (62–64), comparable to what was found in acutely infected mice (65). These results can be explained by the recent finding that urine decreases UPEC expression of type 1 pili (66). Type 1 pili have long been known to play a critical and essential role in establishing cystitis in a murine model of experimental UTI, and vaccination of mice and cynomolgus monkeys with the type 1 pilus-tip adhesin, FimH, protects against experimental cystitis (36, 38, 39, 65, 67–71). Type 1 pili have also been shown to be required for UPEC adherence to human urothelial tissue culture cells, and expression of FimH was required for bacterial adherence to human bladder tissue in situ (39, 56, 72). Lastly, there is strong evidence that FimH has undergone pathoadaptive mutation in UPEC clinical isolates, with several amino-acid residues found to be under positive selection (73–77). Mutation of these residues resulted in reduced virulence in a murine model of cystitis, providing further support that FimH plays an important role in vivo during human UTI (73). Recently, specific pathoadaptive FimH alleles were found to affect the structural conformation and mannose-binding affinity of clinical UPEC isolates, as well as the virulence of the isolates in mouse models of acute and chronic cystitis (78).

FimH mediates UPEC adherence to and invasion of urothelial cells

During experimental bladder infection, the type 1 pilus-associated tip adhesin FimH mediates adherence and invasion of the superficial umbrella cells of the urothelium (27, 65). The specific receptor for type 1 pili appears to vary with the differentiation state of the urothelial cells. In mature superficial umbrella cells, the FimH receptor is the mannosylated-uroplakin protein UPla (58). However, the immature urothelial cells generally used for in vitro studies, such as 5637 bladder transitional carcinoma cells, do not typically express uroplakins on the cell surface, and FimH was found to bind to mannosylated α3 and β1 integrins in vitro (59). Klumpp and colleagues have demonstrated that binding of FimH to the uroplakin receptor complex via UPla leads to the phosphorylation of UPIIIa, the only one of the four major uroplakins with a potential cytoplasmic-signaling domain, resulting in an increase in intracellular calcium and enhanced invasion (79, 80). However, in immature urothelial cells, bacterial invasion subsequent to FimH binding has been reported to involve components of clathrin-coated pits such as...
### TABLE 1 Prevalence and sites of action of selected uropathogenic E. coli (UPEC) virulence factors and their use as candidate vaccine antigens

<table>
<thead>
<tr>
<th>Category</th>
<th>Virulence factor</th>
<th>Prevalence (%)</th>
<th>Animal model</th>
<th>Bladder</th>
<th>Kidneys</th>
<th>References</th>
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<td>36, 39, 66, 70, 300, 407</td>
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<td>N</td>
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<td>VAT/Tsh</td>
<td>54–68</td>
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<td>NR</td>
<td>NR</td>
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<td>Autotransporter (T5SS) adhesins</td>
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<td>70–94</td>
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<td>Conserved</td>
<td>Mice</td>
<td>S</td>
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*Data from references 41, 117, 119, 176, 177, 199–206.
*S, mutant strain was attenuated in single-infection studies.
*C, mutant strain was attenuated in competition studies.
*P, vaccination with virulence factor was protective against challenge infection.
*NR, not reported.
*N, no phenotype.
*(R), attenuation of mutant strain was rescued by complementation.
*L, mutant strain caused less organ damage in single-infection studies.
*IBC, mutant strain formed smaller intracellular-bacterial communities.
*D, mutant strain caused more organ damage in single-infection studies.
clathrin and the cargo-adaptor protein AP-2 (81), caveolae and lipid rafts (82), the action of microtubules (83), and actin rearrangement involving focal-adhesion kinase, phosphotidylinositol-3-kinase, and the Rho GTPases Rac1 and Cdc42 (72, 84, 85). The toxin cytotoxic-necrotizing factor 1 (CNF1) has been reported to enhance UPEC invasion of urothelial cells *in vitro* by constitutively activating Rho GTPases (86), but CNF1 has not been shown to play a clear role *in vivo* (87, 88).

**P** **pili**

The role of P pili in UTI is complex and not fully understood. While P pilus-expressing UPEC are strongly associated with first-time pyelonephritis in children (89, 90), they are less well conserved in women with acute and recurrent UTI, being expressed in only 40% to 50% of isolates, regardless of upper urinary tract involvement (91). This is likely due to the chronic inflammatory changes that occur in the urinary tract of patients with a history of severe or recurrent UTI (33), which may lessen the requirement for P pili in colonizing the kidney. P pili mediate adhesion to Gal-α-1,4-Gal-containing globoseries glycosphingolipids elaborated on the surface of urinary tract epithelial cells (92–95). Patients with upper urinary tract symptoms during UTI mount humoral antibody responses to P pili, indicating that they are expressed during infection (96, 97). Moreover, in humans who are non-secretors of ABO antigens, sialyl galactosyl globosides, which are P pilus receptors, are found more abundantly on the surface of epithelial cells in the kidneys and lower urogenital tract compared to “secretors,” perhaps explaining why some studies have identified the non-secretor status as a significant risk factor for recurrent UTI (98–101). Concordant with these findings, P pilus-expressing UPEC have a greater capacity to bind to vaginal epithelial cells from non-secretors than from secretors (98).

There are at least three alleles of the P pilus tip adhesin PapG, each of which differs in its binding specificity to globosides. Of these, PapGII, which binds the human kidney receptor GbO4, is required for the establishment of pyelonephritis in cynomolgus monkeys (102, 103). Replacement of PapGII with PapGIII, which binds to Forssman antigen (GbO5) and is the predominant PapG allele found in cystitis strains (104), shortened the course of bacteriuria after bladder inoculation of primates and diminished both renal damage and the development of a serum titers against P pili. In a primate model of uncomplicated cystitis, neither PapGII nor PapGIII were required for robust bladder infection (105). However, PapGII conferred a competitive advantage in the primate bladder when co-inoculated with an isogenic strain lacking P pili (PapGIII was not tested in competitive infection). Deletion of P pilus operons, including one expressing PapGII, from a virulent UPEC strain did not affect pathogenesis in a CBA murine model of infection (106). This may be because the UPEC strain used in the study contains alternative adhesins capable of colonizing the kidney epithelium, or perhaps the CBA-mouse model does not reflect the importance of PapGII – GbO4 interactions, since the GbO4 receptor is likely not as highly expressed in the murine kidney due to the presence of a functional Forssman synthetase in non-primate mammals (107). Furthermore, the C3H/HeJ strain, which is closely related to the CBA strain, has been reported to be genetically susceptible to vesicoureteric reflux (108), so the requirement for kidney adhesins may be diminished in this model. In contrast, a mutant UPEC strain lacking PapGII and other putative kidney-colonization factors was defective in colonizing the kidneys of Balb/c mice. Live multiphoton studies of pyelonephritis in rats suggest that P and type 1 pili may work in concert to colonize the renal tubules by facilitating bacterial attachment and biofilm production, respectively (109). Therefore, although P pili are only expressed in about half of all recurrent UTI isolates, drugs that target both type 1 and P pili would likely have broadly protective effects in both the bladder and kidneys.

**Other pili and non-pilus adhesins**

Compared to commensal strains, UPEC have been shown to contain numerous CUP-assembled pilus operons, in addition to those encoding type 1 and P pili. These pilus operons were identified by homology to the four minimum essential components of CUP-assembled fimbrial systems: the major pilin, the pilus adhesin, the outer-membrane usher, and the periplasmic chaperone. However, the functions and receptors of these additional UPEC-associated pili are poorly understood. S and F1C pili and Afa/Dr adhesins are enriched among UPEC, but are less well conserved than type 1 and P pili. They have been demonstrated to impart the capacity to bind to human kidney epithelia in frozen-tissue sections (110, 111) and may play distinct roles in various stages of UTI (112–116). Recently, the Yad pilus was shown to contribute to adherence of UPEC to bladder epithelial cells *in vitro*, but was not required for experimental urinary tract infection in mice, whereas the Ygi pilus conferred a modest competitive advantage in kidney colonization (117). While pili are likely involved in the initial attachment of UPEC to the urinary tract mucosa, the bacteria elaborate numerous other outer-membrane protein...
E. coli UTI89 P pilus

E. faecalis OG1RF Ebp pilus

Tip Adhesin
Major Shaft Subunit
Pilus Base/Terminator
Regulatory Gene
adhesins that may play an important role in disease pathogenesis. Recently, a novel adhesin, TosA, which is secreted by a cognate Type 1 secretion system, was described (118). TosA is found in about 30% of urinary-tract isolates and is expressed during UTI (118–120). However, the role of TosA in UTI is unclear. Although an isogenic UPEC mutant lacking this RTX protein is dramatically impaired in its ability to colonize the urinary tract, the authors of the study were unable to complement the mutant, and vaccination of mice with this protein had no impact on the course of UTI by a TosA-containing UPEC strain (121). Another recently identified adhesin, FdeC, is highly conserved among all E. coli pathotypes and intestinal commensals and is reportedly expressed only upon contact with host cells (122). The presence of FdeC conferred a competitive advantage in colonization of the bladder and kidneys of the mouse model, but vaccination of mice with FdeC antigen only protected against kidney infection, with no effect on bladder infection. Finally, the iron-regulated adhesin Iha has been shown to mediate adherence to bladder epithelial cells and confer a slight, but significant, competitive advantage to UPEC in the mouse model of UTI (123). Thus, despite the considerable progress in our understanding of UPEC pathogenesis over the past 15 years, type 1 and P pili remain the most promising candidate adhesins for drug and vaccine intervention.

Cyclic adenosine monophosphate (AMP) and UPEC expulsion

After internalization, UPEC have been found to reside within Rab27b/CD63/Caveolin-1-positive fusiform vesicles, which resemble secretory lysosomes and are normally involved in regulating the surface area of the apical-plasma membrane. However, UPEC can be expelled by a mechanism that requires Toll-like receptor 4

![FIGURE 2 Models of pilus assembly in Gram-negative and Gram-positive pathogens.](image_url)

**Top panel.** Model of P pilus formation by the chaperone-usher pathway in uropathogenic E. coli. After secretion of pilus subunits into the periplasm via the general Sec machinery, periplasmic chaperones (dark green) serve as folding templates, providing a beta-sheet that enables proper folding of the pilin subunits into immunoglobulin-like domains, but in a non-conical orientation, in a mechanism calleddonor-strand complementation. Assembly and anchoring of the pilus occurs at an outer-membrane pore known as the usher (orange). The pilus-tip adhesin (red) is the first subunit to interact with the usher, via a preferential interaction between the tip adhesin/periplasmic-chaperone complex and the usher N-terminal-periplasmic domain (NTD, light blue), and this interaction initiates assembly by causing a conformational change in the usher that ‘unplugs’ (Plug, dark blue) the pore and displaces the tip-adhesin subunit/chaperone complex to two C-terminal-usher domains, CTD1 (yellow) and CTD2 (purple) (47, 49, 423). The next pilin subunit/chaperone complex then binds to the NTD and if it has an N-terminal extension that is able to complete the immunoglobulin fold of the preceding subunit in a canonical fashion, this provides the free energy to displace the chaperone, in a process called donor-strand exchange, and drive assembly (47–49, 423). In P pili, this occurs repeatedly, incorporating anywhere from hundreds to thousands of PapA major-pilin subunits (green) in the pilus, until PapH (brown) is incorporated into the pilus. PapH is a terminator because it is unable to undergo donor-strand exchange (424). Small-molecule inhibitors (pink) that disrupt pilus assembly (‘pilicides’) or adhesin binding to its receptor (‘pilus-adhesin antagonists’) have been identified (223, 237). **Bottom panel.** Model of sortase-mediated assembly of the endocarditis- and biofilm-associated pilus (Ebp pilus) in E. faecalis (217). Unlike CUP pili in Gram-negative bacteria, sortase-assembled pilus subunits are covalently linked. Pilin subunits are first secreted to the outside of the cell via the general Sec machinery, and are retained in the membrane via a hydrophobic domain within their cell wall-sorting sequence. Sortase C (SrtC, yellow) cleaves the EbpA (red) LPETG sequence, resulting in an EbpA-SrtC chioacyl intermediate that is resolved by the EbpC (green) Lys186 nucleophile. Pilus polymerization occurs when SrtC processes the EbpC LPSTG sequence at the base of a growing, membrane-associated pilus forming a pilus-SrtC intermediate that is resolved by the Lys186 of an incoming EbpC subunit. EbpB (brown) incorporates at the base of a pilus fiber when its Lys179 nucleophile resolves a pilus-SrtC intermediate. Sortase A (SrtA, blue) processing of the EbpB LPKTN sequence leads to eventual incorporation of the mature pilus into the cell wall. Sortase inhibitors (pink) may be useful for disrupting the virulence potential of Gram-positive uropathogens.
(TLR4), cyclic AMP, Rab27b, and caveolin-1 (85, 124–126). Treatment of mice prior to infection with the drug forskolin, which increases cytosolic cyclic AMP, reduces the intracellular bacterial burden in the bladder. Forskolin has also been shown to suppress UPEC-induced inflammation in vitro in primary kidney tubular epithelial cells, and forskolin administration at one hour post-infection reduced acute kidney bacterial burden and inflammation in UPEC-infected C57BL/6 mice (127). Furthermore, TLR4-signaling-incompetent C3H/HeJ mice have higher intracellular bacterial burdens than TLR4-signaling-competent C3H/HeN mice (128). Thus, TLR4-dependent antagonism of invasion and active expulsion of internalized bacteria by urothelial cells is an important early innate defense against acute infection of the bladder, and these expulsion pathways are attractive targets for drug prophylaxis.

**UPEC escape the endocytic vesicle**

Although UPEC may be expelled from host cells after invasion, it is clear that a fraction of invasive bacteria survive within the superficial umbrella cells, eluding expulsion and phagolysosomal death, and escape into the cytoplasm, where several groups have demonstrated that they can replicate rapidly to form IBCs (29, 30, 129–132). Although the mechanism of escape into the cytoplasm is not understood, IBC formation does not typically occur in undifferentiated urothelial cells unless they are treated with either membrane- or actin-destabilizing agents (133, 134). This suggests that the actin network, which is denser in undifferentiated urothelial cells compared to superficial umbrella cells, may restrict bacterial escape from the vesicle and/or proliferation of UPEC within the cytoplasm. It may also be that the difference in FimH receptors in undifferentiated (α3 and β1 integrins) and differentiated (UPIa) urothelial cells results in distinct and divergent UPEC-invasion pathways and intracellular trafficking. While pore-forming toxins have been shown to be required for bacterial escape into the cytoplasm in other infectious-disease model systems, α-hemolysin, a UPEC pore-forming toxin which is expressed in IBCs (135), is not required for vesicular escape, as a mutant strain of UTI89 lacking the α-hemolysin gene forms IBCs equally as well as the wild-type strain (136).

**IBC formation**

Upon escape into the urothelial cell cytoplasm, UPEC replicate quickly to form IBCs, with a doubling time of 30–35 minutes (30). A survey of published studies finds that the number of IBCs detected at 6 hours post-infection (hpi) in the bladders of individual mice ranges from 3 to 700 (median: ~40) after infection of 7–10-week-old C3H/HeN mice with 107 colony-forming units of the UPEC strain UTI89 (28, 71, 73, 137–139). Microscopy studies of mouse bladders after infection with a mixed inoculum of green fluorescent protein-expressing (GFP+) and non-expressing (GFP-) UPEC have demonstrated that IBCs are clonal, originating from a single invasive bacterium (28). As a result, IBC formation appears to constitute a population bottleneck that initially limits bacterial diversity, followed by a rapid expansion of the clonal IBC population. Novel anti-infective drugs that prevent IBC formation may thus target a vital molecular bottleneck, which is thought to be the Achilles’ heel of a pathogen during infection (140, 141). UPEC aggregation into IBCs resembles biofilm formation, as it requires continued type 1 pilus expression after invasion (71) and is accompanied by the production of structural components otherwise associated with UPEC biofilm, such as antigen 43 and a polysaccharide-rich matrix (29). Capsular-synthesis genes also play a role, as a K1 capsule-deficient mutant of the human cystitis isolate UTI89 is markedly deficient in its ability to aggregate and form IBCs (142), and a K2 capsule-deficient mutant of the human pyelonephritis isolate CFT073 is significantly outcompeted by wild type CFT073 in the bladder and kidneys (143). Secreted amyloid fibers (curli) and several other UPEC-autotransporter proteins, including UpaC, UpaG, and UpaH, have been implicated in biofilm growth; however, their role in IBC formation is unknown (144–148). The IBC pathway has been observed in all mouse strains tested, and 15 of 18 human clinical UPEC isolates formed IBCs in experimental infections of C3H/HeN mice, including some isolates without common putative UPEC-virulence factors such as α-hemolysin (149). Those strains unable to form IBCs were also unable to invade the mouse urothelium. UPEC within IBCs are protected from both phagocytosis by polymorphonuclear leukocytes (PMNs) (30) and many antibiotics, particularly the first-line drug trimethoprim-sulfamethoxazole, which has increased efficacy against UTI because it concentrates in the urine but is relatively cell-impermeant (131, 150). A recent study demonstrated that 16 antibiotics capable of killing the virulent cystitis isolate UTI89 in vitro are relatively ineffective in eliminating intracellular bacteria either from bladder epithelial cells in vitro or from bladder tissue during in vivo infection, even though they achieved urine levels far exceeding the minimum-inhibitory concentrations for UTI89 (131). Thus, harboring bacteria that are protected from antibiotics within IBCs or a persistent...
intracellular niche (131, 133, 151) may provide a source of surviving pathogens within the bladder that can cause a relapse (treatment failure) or recurrent cystitis, respectively, once antibiotics are removed.

The IBC pathway occurs in humans and with other Gram-negative uropathogens that express type 1 pili

IBC development is not limited to experimental UPEC infection in mice. Translational studies found evidence of IBCs in 18% of urine sediments from women with recurrent cystitis with UPEC (a rate of detection similar to that seen in the urine of mice acutely infected with UPEC), but never in urine from healthy controls or when the causative agent of the UTI was a Gram-positive organism (31). Other studies have identified IBCs in the urine of children with recurrent UTI (32, 33). Furthermore, other Gram-negative uropathogens that express type 1 pili, such as Klebsiella pneumonia, Enterobacter spp., and Citrobacter freundii, also utilize the IBC pathway (Rosen and Hultgren, unpublished data; 139, 152). Together, these findings suggest that the IBC pathway is an important mechanism for the establishment of UTI in mammalian bladders by Gram-negative uropathogens that express type 1 pili and invade the urothelium. Therefore, the IBC pathway is an important and relevant target for therapeutic intervention.

UPEC dispersal and further IBC formation

Dispersal of UPEC from the IBC is also critical for bacterial persistence. IBC maturation involves a partially understood differentiation program during the first 12–16 hours of experimental infection of the mouse bladder. During this time, the rapidly replicating bacteria first take on a coccoid morphology, become more rod-shaped again as the IBC matures, and then begin to flux away from the IBC. UPEC then emerge out of the dying urothelial cells, often in filamentous form, and colonize and invade neighboring cells, thus initiating a second round of IBC formation (30). Flagellar motility does not appear to be required for UPEC dispersal or initiation of the second round of IBC formation (153). However, deletion of the cell division-inhibitor gene, sulA, disrupts the ability of UTI89 to filament, a property that has been associated with resistance to neutrophil attack. The sulA mutant is also defective in bladder colonization and IBC formation at 24 hpi, but not at 6 hpi, suggesting that UPEC filamentation is necessary for virulence after the first round of IBC formation in the immunocompetent host (154). IBCs are transient, cycling through formation and dispersal primarily during the first 2–3 days of experimental UPEC infection in the immunocompetent mouse (30). However, the immunodeficient-mouse strain C3H/HeJ, which lacks the ability to sense bacterial lipopolysaccharide (LPS), had microscopic evidence of bladder IBCs 4 weeks after experimental infection, indicating that host responses to LPS during infection alter the susceptibility of the bladder urothelium to IBC formation (42). Thus, the IBC pathway is important for the establishment of acute infection in the host, and resembles biofilm formation in the sense that both aggregation and dispersal of UPEC are critical for acute pathogenesis.

Central metabolism and two-component systems

Recently, central-metabolism pathways, such as the tricarboxylic acid cycle, have been shown to be important for acute UPEC virulence and IBC formation in the urinary tract, but not for planktonic growth in urine (155–159). The QseBC two-component system, which is found in many Gram-negative pathogens including UPEC, plays a critical role in regulating virulence-factor expression (160–162). Two-component systems typically consist of an inner membrane sensor kinase and a cytoplasmic response regulator. In response to a stimulus, the sensor kinase regulates by phosphorylation the activation state of the response regulator, thereby regulating gene-expression programs. Dysregulation of the QseBC system by deletion of the sensor kinase QseC causes pleiotropic effects in the bacterial cell, including reduced expression of virulence factors (such as type 1 pili) and reduced virulence and IBC formation in vivo (156, 157). A qseC-deletion mutant forced to express type 1 pili also had an acute virulence defect when in competition with wild-type UPEC, suggesting that the misregulation of additional factors beyond type 1 pili was responsible for the attenuation (163). Surprisingly, we found that the altered virulence-factor regulation in the ΔqseC mutant was due to defects in central metabolism, as two different mutants unable to complete the tricarboxylic acid (TCA) cycle phenocopied the ΔqseC mutant (156). The QseBC system was recently found to have robust and highly sensitive cross-regulation with another two-component system, PmrAB, which is activated by ferric iron and which mediates polymyxin resistance. Addition of ferric iron to UTI89 growth medium induced qseBC expression in a PmrB-dependent manner (164). Other two-component systems also contribute to UPEC virulence. Cpx is an envelope stress-response system known to regulate the expression of
P pili. It was recently shown that deletion of the Cpx system resulted in impaired UPEC colonization of the murine bladder and impaired virulence in zebrafish embryos. Finally, other two-component systems, including PhoP-PhoQ, BarA-UvrY, and KguS-KguR, have been found to contribute to UPEC virulence. Interestingly, constitutive activation of the Pho regulon in CFT073 by inactivation of the phosphate-specific transport system Pst was recently shown to result in a loss of expression of type 1 pili, resulting in significant attenuation in a mouse model of UTI. Therefore, compounds that alter UPEC virulence gene expression or central-metabolism pathways, either directly or by misregulating two-component systems, are potential novel therapeutics.

Metal ions
Iron acquisition is another critical requirement for bacterial virulence. Iron acquisition-associated genes common to all E. coli strains are under strong positive selection in UPEC clinical isolates. UPEC typically have multiple, seemingly redundant iron acquisition systems, and these have been shown to be highly upregulated in the IBC. As many as four siderophores (small-molecule iron chelators) are commonly produced by UPEC strains, and scavenging ferric iron (Fe3+) is thought to be their main function. Among the siderophores, enterobactin is broadly conserved among E. coli strains, while yersiniabactin, salmochelin, and aerobactin-synthesis genes are enriched in UPEC. To prevent microbial iron scavenging, urothelial cells in close proximity to the IBC upregulate genes for the transferrin receptor and for lipocalin 2, host factors that are involved in preventing bacterial acquisition of iron. However, metabolomic studies have found that UPEC clinical isolates preferentially synthesize yersiniabactin and salmochelin, each of which is associated with resistance to the anti-bacterial effects of lipocalin 2. Furthermore, UPEC can scavenge iron from heme, and a deletion mutant lacking the heme transporter ChuA (which is highly expressed in the IBC) forms significantly smaller IBCs in vivo. Siderophore production in E. coli is mediated in part by the small regulatory noncoding RNA RhyB; a recent study found that deleting rhyB in CFT073 reduced siderophore production in vitro and in vivo and reduced bladder and kidney colonization in CBA/J mice.

The salmochelin receptor IroN may have multiple functions, as it has been shown to enhance bacterial invasion of bladder epithelial cells in vitro, and a mutant UPEC strain lacking IroN was attenuated in a mouse model of cystitis. Yersiniabactin also plays a role in sequestering the toxic effects of copper (II) ions, possibly enhancing resistance to phagocyte killing. Interestingly, the broadly conserved siderophore enterobactin actually contributes to copper sensitivity, suggesting that the apparent redundancy of siderophores may actually be a bacterial adaptation to inhibiting different host niches. In mice, the asymptomatic bacteriuria isolate UPEC 83972 outcompeted a mutant strain lacking salmochelin and enterobactin in the urine, bladder, and kidney. In the pyelonephritis isolate CFT073, which does not synthesize yersiniabactin, aerobactin appears to play an important role in bladder fitness, suggesting that these two siderophores may have overlapping functions. Thus, bacterial iron acquisition by multiple systems has been selected for in UPEC, possibly in part due to their role in IBC formation. Their redundancy points to their importance, which may make targeting them with vaccines or therapeutics problematic. Including multiple siderophore-receptor antigens in a single vaccine might possibly overcome this challenge. However, all siderophore receptors require the TonB inner-membrane protein to transduce the energy needed for import. Deletion of TonB from a UPEC strain greatly reduced virulence in the kidney and, to a lesser extent, the bladder during experimental mouse infection. Therefore, targeting TonB with small-molecule inhibitors may be an effective anti-infective strategy.

Modeling the outcomes of acute cystitis
Experimental mouse models of infection have revealed that UPEC are capable of chronic colonization of the urinary bladder in several different ways. In immunocompetent mice, the outcome of cystitis is typically either resolution of acute infection with elimination of bacteriuria, or persistent bacteriuria and chronic cystitis. However, even with resolution of active infection, UPEC are capable of persisting latently within Lamp1+ vesicles inside urothelial cells. These latent reservoirs have been termed the quiescent-intracellular reservoir (QIR) and have the capacity to seed recurrent infections. Treatment of mice with urothelial exfoliations, including protamine sulfate and chitosan, has shown some promise in eliminating this bacterial reservoir from the bladder.

In contrast to a latent reservoir, some mouse strains (including the C3H/HeN and CBA/J strains, which are commonly used for mouse models of UTI) are prone to developing high-titer persistent bacteriuria and chronic cystitis, which appears to last for the life of the animal,
in response to UPEC infection in an infectious dose-dependent fashion (42, 186). C57BL/6 mice also develop chronic cystitis when “superinfected” with two UPEC exposures 24 hours apart (187). Inflammation is most severe during early acute infection in this model and plays a non-productive role, actually contributing to the development of chronic cystitis. This is potentially a very interesting model, as placebo-controlled studies have found that approximately 50% of women remain bacteriuric several weeks after a symptomatic UTI if not treated with antibiotics, despite overall improvement of symptoms (188, 189). Antibiotic therapy readily cures the infection in mice. However, if chronic cystitis is allowed to ensue for at least 7–14 days prior to antibiotic therapy, the mice become highly sensitized to severe, recurrent cystitis upon a second bacterial challenge administered 6 months or more after antibiotic therapy to clear the initial infection (O’Brien, Hannan, and Hultgren, unpublished data; 42). A recent proteomics investigation revealed bladder mucosal remodeling in these so-called “Sensitized” mice that renders the host more susceptible to neutrophil damage as a consequence of inflammation (45). In contrast, mice that are treated with antibiotics after only 1 day of infection, or that spontaneously resolve bacteriuria during the first two weeks of infection, are more resistant to challenge than naïve, age-matched mice. This model may be an invaluable tool not only for understanding host mechanisms of chronic and recurrent UTI, but also for developing therapies and vaccines that combat recurrent UTI (35). For instance, treatment with cyclooxygenase-2 (COX-2) inhibitors prior to challenge infection was found to prevent recurrent UTI in “Sensitized” mice by preventing bladder epithelial transmigration of neutrophils and subsequent mucosal wounding (45). This finding may explain the results of a small study that found that women who received ibuprofen, a non-specific inhibitor of cyclooxygenases 1 and 2, resolved UTI symptoms as quickly as women who received the antibiotic ciprofloxacin did (190).

Ascension to and colonization of the kidneys
The main complication of untreated cystitis in humans is ascension of bacteria in the ureters and colonization of the kidney parenchyma (medulla and cortex), which can lead to marked kidney inflammation with progressive loss of nephron function and even sepsis. Flagella are highly expressed by UPEC in vivo during ascension to the murine kidney (191). However, the contribution of flagella to kidney colonization is unclear, as different mutants impaired in flagellar motility or chemotaxis were not all defective in kidney colonization in competition-infection experiments (153, 192). This may be because the mouse strains used in these studies, C3H/HeN and CBA/J, are genetically susceptible to vesicoureteric reflux (108), and therefore flagellar motility may not be important for ascension in these models. The UPEC pore-forming toxin α-hemolysin is associated with renal damage and scarring (193). At small physiological doses it induces Ca2+ oscillations in renal tubular epithelial cells and thereby potentially enhances ascension and colonization of the ureters and kidney parenchyma by disrupting the normal flow of urine (194). Recently, α-hemolysin was found to induce proinflammatory Caspase-1/Caspase-4-dependent cell death in bladder epithelial cells, resulting in cell exfoliation. UPEC strains overexpressing α-hemolysin were attenuated in acute and chronic infection in mice, suggesting that acute bladder exfoliation is a host defense mechanism (195). Other UPEC toxins, such as Sat, PicU, and Tsh, are also not required for infection, but may contribute to renal pathology (196, 197). Ygi pili, type II- and IV-secretion systems, and multiple iron and heme acquisition components have all been shown to contribute to kidney colonization in animal models (117, 184, 198, 199). Recently, a Toll/interleukin-1 receptor (TIR) domain-containing protein that is secreted by an unknown mechanism, TcpC, was discovered in a subset of UPEC and found to associate directly with MyD88 and TLR4 (200, 201). Loss of function studies in mice indicate that TcpC enhances bacterial virulence by suppressing the early innate immune response to UPEC infection, resulting in higher bacterial burdens in the kidney and more severe kidney pathology over time (202).

Complicated UTI: Enterococcus faecalis
CAUTI
In contrast to the healthy-bladder environment affected by UPEC in uncomplicated UTI, the placement of a urinary catheter effects pathologic changes in the bladder that may contribute to the greater variety of competent uropathogens that infect the catheterized bladder. In addition to causing mechanical damage, urinary catheterization interferes with micturition (urination), a natural impediment to bacterial colonization of the bladder. Even in the absence of infection, bladder catheterization may lead to tissue edema and hyperplasia of the urothelium, in addition to hematuria and alteration of urine composition (203, 204). Probably the most significant factor contributing to infection development is the presence of an abiotic surface in the bladder, the
catheter, which promotes bacterial-biofilm formation and is recognized as an important component of CAUTI. UPEC and enterococci are the two most common isolates from symptomatic CAUTI (14). Enterococci, commensal gut bacteria, have emerged as important human pathogens in the last 40 years, especially in the health care environment. Several aspects of modern medicine have contributed to this recent rise in infections caused by *E. faecium* and *E. faecalis*, the most commonly isolated species. Widespread use of antibiotics has likely selected for enterococci that display intrinsic or acquired resistance to many common classes of antimicrobials. Furthermore, medical devices (such as indwelling urinary catheters) and invasive surgical procedures compromise natural barriers to infection and are being used more frequently in the health care setting. Despite the increasing incidence of enterococcal infection, little is known about the molecular mechanisms these bacteria use to cause disease. However, it is clear that biofilm formation and the elaboration of secreted and surface proteins and organelles contribute significantly to enterococcal pathogenesis.

**The role of secreted and surface structures in enterococcal-biofilm formation**

Biofilm formation is a critical aspect of device-related infections, including enterococcal CAUTI. As extracellular pathogens, enterococci may also rely on growth in the biofilm state to infect host tissues. Thus, putative molecular determinants of enterococcal virulence, identified using a variety of methodologies, are typically examined *in vitro* in assays of biofilm formation. These determinants must be secreted and, in many cases, covalently linked to the cell wall. This latter function is carried out by a group of membrane proteins known as sortases, which recognize conserved-cell wall-sorting sequences (CWSS) on membrane-linked proteins and catalyze the attachment of their extracellular domains to the cell wall. Bacterial proteins well-studied in biofilm assays include enterococcal surface protein (Esp), of unknown function, and gelatinase, a secreted zinc metalloprotease that hydrolyzes gelatin, collagen, and casein (205). Extracellular DNA, autolysin, the housekeeping sortase SrtA, the endocarditis and biofilm-associated (Ebp) pilus, and the Ebp pilus-associated sortase SrtC have also been shown to play a role in biofilm formation (206, 207).

**The role of adherence and biofilm formation in CAUTI**

Many of the same virulence factors involved in biofilm formation, including Ace, Esp, AhrC, Eep, SrtA, Ebp pili, and SrtC, have been shown to play a role in a ureteric-reflux model of ascending pyelonephritis (207–211). However, the contribution of these factors was not large, and robust bladder infection could not be achieved in rodent models of uncomplicated UTI (212). More recently, a more relevant model of foreign-body cystitis has been developed in mice for testing the role of *E. faecalis* virulence factors in CAUTI (213). In this model, the presence of urinary-catheter material in the urinary bladder allows for biofilm formation on the implant and robust, high-titer bladder infection that persists as long as the catheter remains. Transient immunosuppression of mice concurrent with catheter implantation exacerbates *E. faecalis* infection in C57BL/6J mice, suggesting that the presence of the catheter and not the inflammatory response is driving CAUTI (214). Ebp pili, and specifically the metal ion-dependent adhesion site (MIDAS) motif found within the predicted von Willebrand factor A domain of the fibrinogen-binding tip adhesin protein EbpA, are essential for both bladder and implant colonization in this model (215, 216). However, the *in vitro* biofilm determinants autolysin and gelatinase were not required (213). Thus, enterococci utilize Ebp pili to take advantage of the presence of foreign abiotic surfaces and damaged bladder-mucosal barriers to cause CAUTI.

**Sortase-mediated pilus assembly**

Ebp pilus assembly is directed by the action of two different sortases (Fig. 2) (217). These membrane-linked transpeptidase enzymes catalyze the covalent assembly of pilus subunits into a functional pilus that is covalently attached to the peptidoglycan cell wall. Sortases are nearly ubiquitous among Gram-positive organisms and have duplicated and diversified among and within species to perform specific functions (218). Therefore, the development of small molecules to inhibit these enzymes has the potential to yield a wide array of therapeutics that range from broadly anti-Gram positive to specifically targeting virulence processes of single species. A sortase inhibitor targeting Ebp pilus assembly could potentially be beneficial for preventing health care-associated infections, including CAUTI. The Ebp pilins also make attractive vaccine candidates for several reasons, particularly for those individuals in a long-term health care setting (e.g., a nursing home), which strongly increases the risk of acquiring CAUTI. The ebp locus is present in ~95% of *E. faecalis* isolates regardless of source and is highly conserved (219). These proteins are expressed and function on the bacterial cell surface and are important virulence factors. Indeed, immunization
with pilins from Group B streptococci has been shown to be protective in relevant infection models (220, 221). The protective effects of immunization with E. faecalis Ebp pilus components in experimental-disease models are described later in the chapter.

TRANSLATING DISCOVERIES IN PATHOGENESIS: THE DEVELOPMENT OF ANTI-VIRULENCE THERAPIES

Biarylmannose-Derivative FimH Antagonists (Mannosides)

The mannose-binding pocket of FimH is invariant in all strains of uropathogenic E. coli (56), and mutations in these residues disrupt mannose binding and attenuate virulence (56, 73, 75). With information gained from the crystal structures of FimH bound to α-D-mannose and mannose derivatives called mannosides (56, 222–224), we and others have rationally designed biarylmannose-derivative FimH binding inhibitors (223, 225, 226). Using a reiterative process of structure-based design, combinatorial chemistry, and in vitro cell-based screening, lead compounds with excellent cellular potency, low molecular weight, and optimized oral pharmacokinetics were identified. Experimental and pre-clinical translational studies have demonstrated that these optimized mannoside compounds can be given orally to mice either to prevent cystitis or to successfully treat an established bladder infection (150, 225, 227, 228).

Since the mannose-binding pocket of FimH is invariant, mannosides have potent efficacy in preventing acute UTI caused by divergent strains, including the trimethoprim-sulfamethoxazole (TMP-SMZ)-resistant UPEC strain PBC-1 (150) and the multi-drug resistant UPEC strain ST131 (228). Mannoside treatment prior to infection of C3H/HeN mice prevents UPEC invasion of the urothelium and IBC formation, a process that protects UPEC from the effects of many antibiotics (131). Thus, in mannoside-treated mice, UPEC are confined to the bladder-extracellular niche, where they are left exposed to high levels of antibiotics that are commonly used to treat UTI. As a result, although TMP-SMZ alone had no effect on bladder colonization by the resistant strain PBC-1, mannoside-potentiated killing by TMP-SMZ (which concentrates in the urine to levels well above the minimum-inhibitory concentration of PBC-1) to successfully prevent the establishment of UTI by this strain. In a similar way, mannoside also potentiated killing by TMP-SMZ to prevent CAUTI in a foreign-body model of experimental infection (227). Finally, a recent study found that mannoside is also efficacious against the multi-drug-resistant UPEC clone ST131 in acute and chronic experimental infection in C3H/HeN mice. The clinical isolate used in this study was EC958, which is an extended-spectrum β-lactamase strain that is resistant to eight classes of antibiotics, including fluoroquinolones. One prophylactic mannoside dose significantly decreased acute bacterial burdens in the bladder and treatment of chronically infected mice with a single dose of oral mannoside reduced bladder bacterial burdens greater than 1,000-fold (228). If translated to clinical practice, mannosides have tremendous and exciting potential to be an efficacious, safe, and cost-effective new therapy either used in combination with commonly used first-line antibiotics to successfully treat existing uncomplicated cystitis and CAUTI, or used alone as a daily prophylaxis against chronically recurrent cystitis. By reducing the use of antibiotics, and particularly the use of fluoroquinolone antibiotics, in the treatment and prevention of UTI, mannosides could have an immediate and long-term impact on the development of antibiotic resistance in UPEC clinical isolates, which is currently as high as 30% in some studies (21). Furthermore, the unique mechanism of mannoside action, i.e., inhibiting the function of the extracellular FimH pilus tip adhesin by blocking the invariant lectin pocket, likely circumvents the development of resistance due to mutation of the binding pocket, porin mutations, or efflux.

Galabiose PapG Antagonists

In 1982, the efficacy of glycolipids in preventing P pilus binding and in vivo infection was established (229). Determination of the crystal structure of PapGII bound to its receptor, GbO4, elucidated critical details of the adhesin-receptor interaction and allowed for further rational design of galabiose-derived receptor analogs (92, 230, 231). Further studies have identified high-affinity multivalent inhibitors that also inhibit galabiose binding by Streptococcus suis (232–234), though they were not consistently effective in a mouse model of peritonitis. Therapy that combines bioavailable PapG antagonists with mannosides has tremendous potential to treat and prevent UTI.

Inhibitors of the Chaperone-Usher Pathway: Pilicides and Curlicides

Since Gram-negative pili are assembled by the chaperone-usher pathway (CUP), inhibitors of this pathway could be broadly effective against a number of pathogens that require pili for pathogenesis. In collaboration with Fredrik Almqvist, a medicinal chemist, we developed
ring-fused 2-pyridone small-molecule inhibitors that target the CUP periplasmic chaperones (235, 236). We have called these compounds “pilicides.” By screening for inhibitors that prevented type 1 pilus-mediated hemagglutination and in vitro biofilm formation, we identified pilicides with activity not only against type 1 pili, but also against P pili (237). NMR and crystallographic studies of the interaction of a pilicide with the P pilus cognate-periplasmic chaperone, PapD, found that pilicide compounds interacted with a highly conserved region (238) that interacts with the N-terminal domain of the outer-membrane usher (237) (Fig. 2). One highly potent inhibitor of type 1 pilation and biofilm formation in vitro, pilicide ec240, was used for an in vitro transcriptomic and proteomic investigation of pilicide effects on UPEC virulence (239). The ec240 pilicide was found to decrease motility and dysregulate CUP pili, including type 1, P, and S pili.

Curli are amyloid fibers produced by many Enterobacteriaceae that are assembled at the outer membrane by a nucleation pathway of fibrillization (240). By screening for inhibitors of curli-mediated biofilm, we identified 2-pyridone derivatives capable of inhibiting both curli and pili formation. One such “curlicide” that inhibits both type 1 pilus production and curli biogenesis rendered UPEC relatively avirulent in a mouse model of experimental cystitis (144). Further optimization of these pilicide and curlicide compounds has increased their potency dramatically (241–243), and these lead compounds are promising candidates for future drug development.

**Intravesical Therapy with ASB Strain 83972**

Another strategy that is currently under investigation is the use of an avirulent asymptomatic bacteriuria (ASB) strain, 83972, which has adapted for long-term colonization of the human urinary tract without causing significant symptoms or pathology, as a therapy for recurrent UTI (244). Although ASB strain 83972 is discussed in more detail in “Asymptomatic Bacteriuria and Bacterial Interference” by Nicolle et al., we will briefly discuss its therapeutic potential here. Strain 83972 has lost the capacity to express type 1, P, and F1C pili, and can outcompete UPEC strains in human urine whether growing planktonically or in a biofilm (245–247). Therefore, it is hypothesized that colonization of the bladder by ASB strain 83972 prevents virulent UPEC strains from colonizing the urinary tract, thereby preventing recurrent symptomatic UTI. This therapy is currently undergoing clinical trials and has shown promise in “at-risk” populations, such as those with incomplete bladder emptying or neurogenic bladder from a spinal-cord injury (248–250). The development of urinary symptoms after 83972 inoculation is rare, and apparently not caused by bacterial reversion to virulence (251). Interestingly, intravesical administration of 83972 to mice with acute UPEC UTI was found to reduce visceral pain, suggesting that 83972 may be an effective treatment for UTI symptoms (252). A variation of this approach was recently employed by Schembri and colleagues. The PapGI receptor, GbO4, is also the receptor for Shiga toxin. Adapting existing technology, Schembri and colleagues engineered a strain of 83972 to synthesize a galabiose analog that is linked to LPS on the surface of the cell (253, 254). This strain is able to inhibit binding of PapGI-expressing UPEC to kidney epithelial cells, and when co-inoculated into the mouse urinary tract with virulent UPEC, the galabiose-expressing strain significantly reduces the UPEC-bacterial load in the urine compared to the wild-type 83972 strain. Conversely, others have transformed strain 83972 with a plasmid expressing type 1 pili and have demonstrated that this new strain forms better biofilm on urinary catheters and as a result is more efficacious in preventing their colonization by enterococci (255, 256).

**Nutraceuticals**

So-called “nutraceuticals” are foods or food products that are thought to provide medical benefits and are often sold in a medicinal form. Since use of these products typically does not require regulatory authority approval (e.g., approval by the Food and Drug Administration in the United States), the efficacy of these compounds is often based merely on anecdotal reports. Two common nutraceuticals that have been investigated for preventing recurrent UTI are probiotic *Lactobacillus* preparations and cranberry products. These products may be advantageous because they are generally safe and readily available. However, there exists only limited evidence for their effectiveness.

**Probiotics**

Clinical evidence suggests that the vagina and periurethral area, which is normally colonized by *Lactobacillus* spp. in healthy women, can act as a UPEC reservoir that could potentially seed recurrent infections. Women with recurrent UTI are more likely to have vaginal or periurethral UPEC colonization than women without recurrent UTI (257), and periurethral-UPEC carriage dramatically increases in the days prior to a recurrent episode (258). Vaginal *Lactobacillus* suppositories might help clear this UPEC reservoir, preventing...
recurrences. However, probiotic therapy with *Lactobacillus* spp. has had mixed results and a recent meta-analysis found no evidence for efficacy (259–261). Clinical trials for *Lactobacillus* prophylaxis for infectious diseases of the urogenital tract, including UTI, are ongoing both in the United States and abroad. A Phase I trial to test the safety of an *L. crispatus* vaginal suppository (LACTIN-V, Osel, Mountain View, California) found that no severe adverse events occurred, although seven women (out of 15) developed asymptomatic pyuria (262). In the randomized, placebo-controlled Phase II trial, women with a history of recurrent UTI received antibiotic therapy for acute UTI, and then either LACTIN-V or placebo. The probiotic was protective, with recurrent UTI occurring in 15% of women receiving LACTIN-V and 27% of women receiving placebo (263). The mechanism of *Lactobacillus*-mediated protection from UTI is not clear, and may involve hydrogen peroxide production. Some *Lactobacillus* strains produce surfactants and anti-adhesive molecules (260). *Lactobacillus acidophilus* surfactant was shown to inhibit initial deposition rates and adhesion numbers for several uropathogens, including *E. coli*, *E. faecalis*, and *Proteus mirabilis* (264). This raises the possibility that some *Lactobacillus* strains might be more effective than others at preventing UTI.

**Cranberry products**

Cranberry products are a common folk-remedy for preventing recurrent UTI, but the efficacy of cranberry products in UTI prophylaxis is largely unproven. Cranberries contain two compounds that have been shown to inhibit UPEC adherence to eukaryotic cells *in vitro*: fructose (found in all fruits), which weakly blocks type I pilus-mediated binding, and A-type proanthocyanidins, which have been shown to block P pilus-mediated binding (265–268). UPEC that was grown in human urine collected after consumption of cranberry juice had significantly reduced adherence to human red blood cells, resin beads coated with P-receptor oligosaccharides, and urothelial bladder cells compared to UPEC grown in normal urine (269, 270). Cranberry products are also less expensive and better tolerated than antibiotics, and thus are an intriguing candidate for UTI prophylaxis. However, the literature regarding the clinical efficacy of cranberry-prophylactic therapy remains inconclusive. For example, a recent meta-analysis of 10 randomized clinical trials found some benefit for women with recurrent UTI, but studies of protection in elderly or catheterized patients are lacking (271). A 2009 randomized controlled trial found that cranberry extract had similar efficacy to low-dose trimethoprim for preventing recurrent UTI in older women (272), while two randomized controlled clinical trials showed no significant effect of cranberry on UTI recurrence in adult pre-menopausal women (273, 274). Comparisons of these different studies may be confounded by differences in type of cranberry product consumed (e.g., juice vs. extract) and dosage regimens. Therefore, more studies are necessary to determine the effectiveness of cranberry products for preventing recurrent UTI.

**Estrogen Therapy**

Another therapeutic approach is the intravaginal application of estrogen. In a controlled trial of post-menopausal women with recurrent UTI, intravaginal application of a topical estriol cream significantly reduced the incidence of recurrence (275). The efficacy was attributed to a restoration of low vaginal pH and vaginal *Lactobacillus* colonization. In contrast, several studies have indicated that systemic estrogen-replacement therapy is not protective against recurrent UTI (276). A recent study found that vaginal estradiol therapy in post-menopausal women altered the expression of antimicrobial peptides and cell-junction proteins in epithelial cells isolated from voided urine, suggesting that estrogen therapy modulates the mucosal barrier of the lower urinary tract (277). In support of these translational findings, several studies of experimental UTI in ovariectomized mice have demonstrated that altering estrogen levels has profound effects on UTI pathogenesis (277–279). Thus, vaginal estrogen therapy remains a safe and viable therapeutic option for post-menopausal women suffering from recurrent UTI.

**Possible Applications to CAUTI**

In contrast to uncomplicated UTI, CAUTI is dependent upon the presence of a foreign body, and removal of the catheter is often curative. Although it is not clear whether the biofilm forms first on the catheter, which in turn allows colonization of the damaged bladder mucosa, or vice versa, biofilm formation is clearly strongly associated with CAUTI, and the most common CAUTI pathogens, UPEC and *E. faecalis*, are good at making biofilm. The use of antibiotic- or silver-impregnated catheters has shown some efficacy in reducing the occurrence of bacteriuria in catheterized patients, but it is unclear whether they lower the rate of symptomatic CAUTI and associated complications (17). For those individuals who require long-term catheter placement, and particularly those with spinal-cord injuries or in...
nursing homes, the additional use of anti-infective drugs such as mannosides and sortase inhibitors in combination may help to prevent a large percentage of CAUTI, as well as infections of other implants. Furthermore, vaccines that target UPEC, E. faecalis, and Proteus spp. may also benefit these patients, reducing the incidence of CAUTI and potentially also the risk for bloodstream infections.

**UTI VACCINES**

**Historical Perspective**

Vaccines have been used against UTI for more than a century, though initially their intended purpose was therapeutic rather than prophylactic. In 1909, two case reports of pregnant women with pyelonephritis described significant clinical improvement after therapeutic systemic vaccination with E. coli (previously known as Bacillus coli) isolated from the urinary tract of the same patient (280, 281). Despite these anecdotal reports, by the 1920s therapeutic vaccination against UTI was largely seen as ineffective. A survey of a thousand American physicians reported little to no use of vaccine therapy for cystitis and pyelonephritis (282), and it was said that “the day of extravagant expectations from vaccine therapy [for UTI] is for the moment past (283).” As our understanding of vaccines advanced, UTI vaccines as a prophylactic therapy (i.e., “clinical” vaccines) reemerged as a topic of interest by the 1950s (20, 284), and have been the focus of much research, refinement, and testing in animals for the past 60 years. Efficacy in animals has been shown for UTI vaccines in every classical category: attenuated, inactivated, subunit, toxoid, and conjugate (Table 2). However, few modern vaccines have been tested in humans, and only one is currently commercially available.

**Challenges in Developing UTI Vaccines**

Clinical and technical challenges associated with developing a clinical UTI vaccine include our lack of understanding of the mechanisms that induce protective immunity in the urinary tract; the diverse patient sub-populations that would benefit from a vaccine; the heterogeneity of UPEC strains, which complicates the choice of the best target antigens; the route of administration (mucosal vs. systemic); and the choice of adjuvant, if needed. Furthermore, experimental conditions tightly control for genetics (often using inbred animals) and environment, whereas the human population is outbred and regularly encounters a broad diversity of environmental variables that could affect the mucosal immune system. When testing UTI vaccines in humans, care must be taken to assess potential confounding variables. It is known that many common vaccines (e.g., influenza, pneumococcal, and zoster vaccines) do not induce optimal immune responses in a large portion of infants and the elderly (285). In addition, hysterectomy may decrease the effectiveness of vaginal-mucosal vaccines, as the cervix has been shown to be “the major inductive and effector site for cell-mediated immunity in the lower female genital tract (286).”

**Recurrent UTI and protective immunity**

The high frequency of recurrent UTI indicates that many patients are unable to mount an effective adaptive-immune response that prevents re-infection. The reasons for this are unknown. One possibility is that uropathogens may mask themselves from and/or directly suppress the immune system. For example, the attenuated UTI vaccine NU14ΔwaaL stimulates the immune system much more than wild-type NU14, which requires waaL O antigen ligase for host-immune suppression (287). Recent studies have found evidence that UPEC infection can promote a tolerogenic bladder environment, suggesting that UPEC exposure can both promote and prevent inflammation (288, 289). Alternatively, the body may downregulate its own immune response to uropathogens in order to maintain the integrity of the mucosal barrier (42, 290). Animal models of recurrent UTI have begun to shed light on the mechanisms of adaptive immunity in response to urinary tract infection. C57BL/6J mice that are repeatedly infected with UPEC become more resistant to experimental UTI (44). However, in C3H/HeN mice, which are genetically susceptible to chronic cystitis in an infectious dose-dependent manner, the outcome of the first infection in naive mice determines susceptibility to subsequent UTI (42, 45). Mice that spontaneously resolve the first episode of cystitis are resistant to bacterial challenge, but those that develop chronic infection upon the first infection are highly susceptible to severe, recurrent infection after antibiotic therapy, even when challenged by less virulent strains that do not cause severe infection in naive mice (O’Brien, Hannan, and Hultgren, unpublished data; 42). Understanding this puzzle will be critical in order to rationally design UTI vaccines with maximal therapeutic efficacy.

**Who should receive an UTI vaccine?**

Cystitis accounts for 90% of all UTI and recurs at high frequency, with 20% to 30% of women experiencing a recurrence within 3–4 months (7). These women
are excellent candidates to receive a cystitis vaccine to lower the rate and severity of subsequent recurrences. The target population for a pyelonephritis vaccine is more limited. Children with vesicoureteral reflux (VUR), diabetics, and newly pregnant women or women of child-bearing age (pre-natal) might benefit from a pyelonephritis vaccine. Both children with VUR and diabetics are at higher risk for developing pyelonephritis (291, 292) and therefore might benefit from vaccination. Importantly, the use of antibiotic prophylaxis to reduce the frequency and severity of VUR-associated UTI in children is controversial among clinicians, and a systematic review of 20 randomized, controlled trials found no clear benefit for antibiotic prophylaxis (293, 294). Thus, new treatment strategies, such as vaccination, are needed for this patient population. Bacteriuria that progresses to pyelonephritis during pregnancy is associated with poor outcomes, including perinatal death, low birth weight, prematurity, and preterm low birth weight (295, 296). However, aggressive treatment of asymptomatic bacteriuria in pregnancy ensures that only a small percentage of pregnant women progress to pyelonephritis (297). Pregnant women with asymptomatic bacteriuria could potentially benefit from a pyelonephritis vaccine, if it were extremely effective and safe for both the mother and fetus. Systemic vaccination that induces immunoglobulin (Ig)G antibodies could also have the potential benefit of conferring passive immunity to the developing fetus, which could protect newborns during their first year or so of life. Indeed, a systemic P pilus-subunit vaccine administered to pregnant rhesus monkeys protected the newborns from pyelonephritis and induced a significant antigen-specific IgG response in the sera of both the mothers and newborns (298).

Choice of immunogen(s)

Effective UTI vaccines should target one or more surface-exposed bacterial structures that are either uniformly expressed by the uropathogen in the host or are expressed during critical stages of infection (Fig. 1). UTI vaccines in development can generally be categorized into two broad categories: “whole-agent” or “whole-cell” vaccines, which include whole bacteria (either live-attenuated or inactivated) and bacterial lysates, and “specific-antigen” vaccines, which include one or more antigens (subunit, toxoid, or conjugate vaccines). Although the majority of vaccines currently licensed in the United States are whole-agent vaccines, most of these target viral pathogens. Among the eight bacterial pathogens for which there are licensed vaccines (compared to 16 vaccines targeting viruses), only three are targeted with whole-cell vaccines. This is because, for bacterial pathogens, specific-antigen vaccines are generally much safer than whole bacterial cell vaccines, particularly when vaccinating systemically with Gram-negative bacteria, which can lead to endotoxemia. However, in the absence of whole organisms, isolated antigens typically do not elicit robust or long-lasting immune responses, and thus must be administered with adjuvants to increase the inflammatory response to the antigen, thereby enhancing immunogenicity (299).

Specific-antigen vaccines can only be as good as the antigen(s) selected. The process of selecting the best antigen(s) represents a critical and formidable challenge early in vaccine development. An ideal vaccine target antigen would be highly and broadly expressed and would be required either for the initiation and/or maintenance of infection or for disease symptoms. For example, toxoid (inactivated toxin) vaccines are highly successful against diseases in which the toxin itself is the main cause of disease, such as tetanus and diphtheria. The development of subunit (protein antigen), conjugated (carbohydrate antigen conjugated to antigenic protein), and DNA (protein expressed from a DNA vector inside the host) vaccines has been the focus of much research, but has been met with limited success. The challenge is that bacterial infections are often much more complex than viral infections, involving multiple host niches and more antigenic variation. Additionally, bacteria have evolved to have redundant virulence mechanisms, a fact highlighted by the multiple adhesins and iron-acquisition systems in many UPEC isolates (53–55). Furthermore, mechanistic studies of bacterial pathogenesis must be carried out in human cell lines or in animal models, and may not accurately reflect the requirement for vaccine targets to initiate infection and cause human disease. Finally, with the exception of the iron-acquisition factors and type 1 pili, the majority of putative UPEC-virulence factors that have been described are found in about 50% or fewer of all isolates (Table 1), and thus would only be useful in a multivalent vaccine.

Once a candidate antigen is chosen, it must be tested in animal-infection models, but the results of such efficacy studies may be difficult to parse. Several potential confounders that can vary among studies must be assessed. Among vaccines comprising whole bacteria, outer-membrane vesicle, and membrane-protein extract preparations, the method of bacterial preparation can have profound effects on the efficacy of the vaccine, as the antigens present will vary with the culture conditions. For example, UPEC grown statically at 37°C in
<table>
<thead>
<tr>
<th>Multi-species vaccines</th>
<th>Vaccine/antigen</th>
<th>Animal model$^a$</th>
<th>Human trials</th>
<th>Site of protection</th>
<th>Route$^b$</th>
<th>Adjuvant$^c$</th>
<th>Antibody responses</th>
<th>References</th>
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<tbody>
<tr>
<td>Whole cell: inactivated</td>
<td>SolcoUrovac: E. coli strains and 1 strain each of P. mirabilis, M. morganii, E. faecalis and K. pneumoniae</td>
<td>M</td>
<td>Trial in children and women Phase I &amp; II clinical trials</td>
<td>Rodent bladder and kidney Human bladder</td>
<td>IM V</td>
<td>None</td>
<td>Increased total and antigen-specific urinary IgG &amp; IgA in mice Increased urinary sIgA Increased total IgG and IgA in vaginal wash and urine</td>
<td>326, 329–335, 419, 420</td>
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<td>Whole cell: inactivated</td>
<td>StroVac: E. coli, P. mirabilis, M. morganii, K. pneumoniae and E. faecalis</td>
<td>None reported in the literature</td>
<td>Distributed by Strathmann GmbH (Hamburg) in parts of Europe, Latin America, and the Middle East</td>
<td>Bladder</td>
<td>IM</td>
<td>Aluminum phosphate N Rd</td>
<td>StroVac Product Insert</td>
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<thead>
<tr>
<th>UPEC-targeted vaccines</th>
<th>Vaccine/antigen</th>
<th>Animal model$^a$</th>
<th>Human trials</th>
<th>Site of protection</th>
<th>Route$^b$</th>
<th>Adjuvant$^c$</th>
<th>Antibody responses</th>
<th>References</th>
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<tr>
<td>Whole cell: inactivated</td>
<td>Heat- or formalin-killed E. coli</td>
<td>R</td>
<td>Led to the development of SolcoUrovac/StroVac</td>
<td>Bladder, kidney</td>
<td>IV IP B IM V</td>
<td>Freund’s</td>
<td>Anti-E. coli IgG and IgA in urine</td>
<td>20, 318, 319, 321–324</td>
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<td>NU14 $\Delta$waal.</td>
<td>M</td>
<td>NR</td>
<td>Bladder</td>
<td></td>
<td>B</td>
<td>None</td>
<td>NR</td>
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<tr>
<td>Whole cell: attenuated</td>
<td>S. enterica serovar Typhimurium expressing 5 pili</td>
<td>R</td>
<td>NR</td>
<td>Kidney</td>
<td>Oral</td>
<td>None</td>
<td>Antigen-specific IgG and IgA in serum</td>
<td>370</td>
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<tr>
<td>Whole cell: attenuated</td>
<td>P pili: purified recombinant synthetic peptides</td>
<td>M</td>
<td>NR</td>
<td>Kidney</td>
<td>ID IM IV SQ LN</td>
<td>Freund’s</td>
<td>Anti-P IgG in serum and urine anti-P pilus IgM in serum</td>
<td>37, 298, 357, 365–368</td>
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<tr>
<td>Specific antigen: subunit</td>
<td>FimH adhesin:</td>
<td>M</td>
<td>Phase I and II clinical trials performed by MedImmune</td>
<td>Bladder</td>
<td>SQ IM IN</td>
<td>Systemic: Freund’s MF59</td>
<td>Anti-FimH IgG in urine and serum Anti-FimH IgA in vaginal washes</td>
<td>38, 39, 300</td>
</tr>
<tr>
<td>Specific antigen: subunit</td>
<td>FimCH</td>
<td>NHP</td>
<td>License recently acquired by Sequoia Sciences</td>
<td></td>
<td></td>
<td>Mucosal: CpG</td>
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<td>FimH truncate</td>
<td>M</td>
<td></td>
<td></td>
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<tr>
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<td>PapDG adhesin</td>
<td>NHP</td>
<td>NR</td>
<td>Kidney</td>
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<td>Aluminum phosphate</td>
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<td>IroN siderophore receptor</td>
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<td>NR</td>
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<td>FyuA siderophore receptor</td>
<td>M</td>
<td>NR</td>
<td>Kidney</td>
<td>IN</td>
<td>CT</td>
<td>Anti-FyuA IgG in serum (correlated with renal bacterial load) and IgA in urine</td>
<td>374</td>
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<tr>
<td>Specific antigen: subunit</td>
<td>Dr adhesins</td>
<td>M</td>
<td>NR</td>
<td>Reduced mortality</td>
<td>NR</td>
<td>Freund’s</td>
<td>Anti-Dr IgG in serum, but no significant effect on colonization</td>
<td>371</td>
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<tr>
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<td>S pili</td>
<td>R</td>
<td>NR</td>
<td>Kidney</td>
<td>SQ</td>
<td>None</td>
<td>Anti-S IgG in serum</td>
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<td>Specific antigen: toxoid</td>
<td>α-hemolysin</td>
<td>M</td>
<td>NR</td>
<td>Kidney</td>
<td>IM</td>
<td>Freund’s</td>
<td>Anti-α-hemolysin IgG in serum</td>
<td>193</td>
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<td>Specific antigen: conjugated</td>
<td>O antigen</td>
<td>RNHP</td>
<td>NR</td>
<td>Bladder, kidney</td>
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<td>Freund’s</td>
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<td>K13 antigen</td>
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<td>R</td>
<td>Kidney</td>
<td>DT</td>
<td>Freund’s</td>
<td>Anti-K13 IgG and IgM in serum</td>
<td>344, 345</td>
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<tr>
<td>Specific antigen: (multi-epitope) subunit</td>
<td>IreA, Hma and IutA iron receptors</td>
<td>M</td>
<td>NR</td>
<td>Bladder, kidney</td>
<td>IN</td>
<td>CT</td>
<td>Antigen-specific IgG and IgM in serum and IgA in urine</td>
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**Proteus-targeted vaccines**

<table>
<thead>
<tr>
<th>Whole cell: inactivated</th>
<th>Heat-killed P. mirabilis</th>
<th>R</th>
<th>NR</th>
<th>Kidney</th>
<th>SQ</th>
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<th>NR</th>
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<tbody>
<tr>
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<td>L. lactis MrpA</td>
<td>M</td>
<td>NR</td>
<td>Kidney</td>
<td>IN</td>
<td>None</td>
<td>Anti-MrpA IgG and IgA in serum</td>
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</tr>
<tr>
<td>Specific antigen: subunit</td>
<td>MrpH adhesin</td>
<td>M</td>
<td>NR</td>
<td>Bladder, kidney</td>
<td>IN, V</td>
<td>CT</td>
<td>High levels of IgG in serum, but no correlation with protection</td>
<td>383</td>
</tr>
<tr>
<td>Specific antigen: subunit</td>
<td>MrpA pilus subunit</td>
<td>M</td>
<td>NR</td>
<td>Bladder, kidney</td>
<td>IN, SQ, V</td>
<td>Freund’s CT</td>
<td>Anti-MrpA IgG in serum and urine</td>
<td>385, 386, 388</td>
</tr>
<tr>
<td>Specific antigen: subunit</td>
<td>UcaA adhesin</td>
<td>M</td>
<td>NR</td>
<td>Bladder, kidney</td>
<td>SQ</td>
<td>Freund’s</td>
<td>Anti-UcaA IgG in serum, but no correlation with protection</td>
<td>385</td>
</tr>
<tr>
<td>Single antigen: Toxoid</td>
<td>Proteus toxin agglutinin (Pta)</td>
<td>M</td>
<td>NR</td>
<td>Kidney, spleen</td>
<td>IN</td>
<td>CT</td>
<td>Anti-Pta IgG in the serum</td>
<td>381</td>
</tr>
</tbody>
</table>

*Animal models: M, mice; NHP, non-human primates; R, rats; Rb, rabbits.*
*Routes of delivery: B, intravesical; ID, intradermal; IM, intramuscular; IN, intranasal; IP, intraperitoneal; IV, intravenous; LN, intra-lymph node; SQ, subcutaneous; V, vaginal instillation; VS, vaginal suppositories.*
*Adjuvants: CT, cholera toxin; DT, diphtheria toxoid.*
*NR, not reported.*
LB liquid media will predominantly express type 1 pili, whereas UPEC cultured on tryptic soy agar plates at 37°C will express P pili. This bias can be overcome with live-attenuated vaccines, which can replicate, mimic the natural route of infection, and change their gene expression accordingly once introduced into the host. Other variables in testing UTI vaccine efficacy in animal models include the animal-infection model and choice of uropathogen used for bacterial challenge, the culture conditions of the challenge-bacterial inoculum, and, as we discuss below, the vaccine-inoculation regimen, including the route and frequency of immunization and choice of adjuvant.

**Route of administration**

Both mucosal (vaginal and intranasal) and systemic UTI vaccines have been effective in animal models. In general, mucosal vaccines elicit both IgA and IgG responses and systemic vaccines elicit only IgG responses. Since IgA is thought to be protective against intimate and invasive infection of the gut, and is found in high concentrations at mucosal sites, it has been traditionally assumed that IgA is the most effective means of inducing mucosal immunity. However, one group compared systemic and mucosal routes of vaccination with the FimH adhesin and found that although only the mucosal route induced elevated levels of vaginal wash and urine antigen-specific IgA in mice, both vaccine delivery routes resulted in similar serum and urine antigen-specific IgG responses and protection against experimental UTI (300). It is possible that this finding may be explained by the experimental model, in which UPEC were instilled directly into the bladder, thus avoiding the initial stages of periurethral colonization and urethral ascension, where IgA may be more important for protection. However, systemic vaccination and serum IgG responses have provided protection against other mucosal pathogens, such as rotavirus and human papillomavirus (301, 302). Therefore, antigen-specific IgG may be equally or more important than IgA for host defense at some mucosal surfaces. Also, data from experimental UPEC infection in mice suggest that the urogenital mucosa may become “sensitized” to uropathogens subsequent to an initial chronic bladder infection (42, 45). In these individuals, a vaginal-mucosal route of vaccine delivery may exacerbate this sensitization.

**Vaccine adjuvants**

The choice of adjuvant can be critical for adequate stimulation of the immune system, but relatively little is known about how adjuvants work (303), and only a few are approved for use in humans. In order to be approved for use in clinical vaccines, adjuvants must have low toxicity. Adjuvants currently approved for use in humans include aluminum salts (e.g., alum), the squalene-based MF59, the LPS-derived monophosphoryl lipid A (MPL), and liposomes (304). The aluminum-based adjuvants aluminum phosphate and aluminum hydroxide are commonly used in systemic vaccines in humans. The specific functions of aluminum adjuvants continue to be debated, but in general, it is accepted that they form a depot at the injection site, allowing for efficient uptake of antigen by antigen-presenting cells (APCs). They also stimulate the immune system by inducing eosinophilia and activating complement and macrophages (305). Of note, alum is a poor stimulator of cellular (TH1) immune responses (306). MF59 is a squalene-based oil-in-water emulsion. After intramuscular injection, the squalene-emulsion droplets are internalized by dendritic cells and enhance antigen presentation (307). Gene-expression analysis of mouse muscle found that, compared to alum and the TLR9 agonist CpG, MF59 induces more changes in gene expression and is a stronger inducer of genes involved with cytokine responses, leukocyte migration, and antigen presentation (308). As a result, MF59 elicited a more rapid influx of myeloid cells to the site of injection. Monophosphoryl lipid A has been modified to reduce its toxicity, while retaining its ability to induce inflammation. It is an agonist for TLR4, although it is unclear whether this agonism is the main cause of its efficacy as an adjuvant. Due to its hydrophobicity, it has a strong propensity to aggregate into microparticulates that are potent activators of the NLRP3 inflammasome (309). Liposomes are thought to enhance immunogenicity, both by enhancing phagocytosis by APCs and by enabling direct cytoplasmic delivery of antigens by membrane fusion (310). Recently, combinations of the above adjuvants have been the subject of much research. In particular, MPL in liposomes has shown great promise, as liposomes have the dual benefit of masking the residual toxicity of MPL while enhancing its potency as an adjuvant (310).

**Mast cell-derived adjuvants**

Mast cells are important players in the bladder innate-immune response. Not only have they been implicated in early defense against UTI in mice, but mast cell-derived factors play an important role in directing the adaptive-immune response (311–313). Mast cell-derived adjuvants are an interesting recent development in vaccinology. Nasal instillation of vaccine antigens...
along with small-molecule mast-cell activators resulted in antigen-specific serum IgG and mucosal (nasal, vaginal, fecal) IgA responses that correlated with increased dendritic cell and lymphocyte recruitment to the lymph nodes (312). Recently, Abraham and colleagues described the synthesis of submicrometer particles that model mast-cell granules and showed their successful use as an adjuvant in a mouse model of influenza (314). An advantage of these particles is that they can be engineered to contain particular cytokines in order to skew the adaptive-immune response. To the best of our knowledge, mast cell-related adjuvants have not yet been tested in UTI vaccines. However, they are an intriguing candidate for further study.

Innate Immunity and the Rise of Systems Vaccinology

In the past, vaccine development was most often a hit-or-miss venture, with little understanding of why some vaccines are efficacious and others are not. To a certain extent this is still true today, but in recent years, vaccinologists and immunologists have begun to understand the role of the innate-immune system in vaccine efficacy. The innate-immune system relies on pattern-recognition receptors (PRRs) expressed by innate-immune cells in order to detect pathogen-associated molecular patterns (PAMPs). An important category of PRRs are the Toll-like receptors (TLRs), which can detect molecular patterns commonly found in bacteria, viruses, fungi, and parasites; C-type lectins and nucleotide oligomerization domain (NOD)-like receptors are also important innate-sensing receptors. Signaling by PRRs on innate-immune cells can trigger an adaptive-immune response that differs based on the PRR and the dendritic-cell subtype (313). Recently, systems biology approaches have been used to assess the effects of vaccination on the immune system, with a particular focus on the early innate response, which can predict vaccine immunogenicity (285). This “systems-vaccinology” approach was used by Pulendran and colleagues to investigate changes in the human immune system after vaccination with a live-attenuated yellow fever vaccine. By performing multiplex-cytokine assays and microarrays with blood collected at baseline and at different time-points post-vaccination, the authors were able to characterize a “molecular signature” involving the innate sensing of viruses that very accurately predicted the development of antiviral immunity (316). The same group subsequently used systems vaccinology to compare immune responses among vaccines for yellow fever, influenza, and meningococcus, revealing distinct transcriptional responses that correlated with antibody responses specific to each vaccine (317). Systems-vaccinology approaches may be useful for assessing the immunological profiles of UTI vaccines, predicting efficacy in vaccinated individuals, and determining the best adjuvant for a given vaccine. It is interesting to note that the mechanism of protection of the only commercially available UTI vaccine, StroVac, is currently unknown. Systems vaccinology may be the key to elucidating the efficacy of this and other UTI vaccines.

Whole-Cell Vaccines in Development

Whole-cell vaccines have been among the most successful vaccines developed to date. Indeed, the only UTI vaccine currently available for use in humans is the polyvalent-inactivated whole-cell vaccine StroVac. Vaccines comprising whole uropathogens, whether attenuated or inactivated, expose the host to a variety of virulence factors. These preparations may or may not include, depending upon how the preparation is grown and processed, pili and other adhesins, outer-membrane proteins, toxins such as hemolysin and CNF1, siderophore receptors, flagellin, and LPS (Fig. 1). Of all vaccine types, live-attenuated vaccines have the potential to most closely mimic natural infections, and thus elicit strong immune responses. However, they cannot be given to immunocompromised patients, and there may be a risk of reversion to virulence in healthy individuals. Inactivated vaccines are generally safer than live ones, but this can be accompanied by the tradeoff of eliciting a weaker immune response than live-attenuated vaccines.

Inactivated vaccines

Inactivated E. coli vaccines have been investigated since at least the 1950s and have been found to protect animals from UTI. An early UTI vaccine consisted of intravenously injected, heat-killed E. coli, and was protective against pyelonephritis in rabbits (20). In the 1970s, systemic vaccination with heat-killed or formalin-killed E. coli strains grown in trypticase soy broth (TSB, which induces P pili expression) protected rats from retrograde E. coli pyelonephritis (318) and ascending UTI (319), but did not protect rabbits against hematogenous pyelonephritis (320). Rats that were vaccinated by intravesical instillation of formalin-killed E. coli were protected against ascending UTI and resolved UTI faster than non-vaccinated controls (319, 321). To the best of our knowledge, vaginal-mucosal immunization with an inactivated-UTI vaccine was first published in 1982, when vaginal instillation with formalin-killed E. coli
protected rats from cystitis (322). In a later study, vaginally instilled, but not systemically injected, formalin-killed E. coli inhibited bacterial adhesion to rat bladder mucosae (323). In 1987, vaginal immunization with formalin-killed E. coli protected cynomolgus monkeys from cystitis (324), and in 1995, intramuscular injection of formalin-killed E. coli protected monkeys from pyelonephritis-associated renal scarring (325).

Urovac and StroVac

With several decades of research showing the efficacy of inactivated-UTI vaccines in animals, Solco Basel Co. developed SolcoUrovac for use in humans. SolcoUrovac was a polyvalent whole-cell vaccine consisting of 10 strains of heat-killed uropathogens: six from E. coli of different serotypes and one each from Klebsiella pneumoniae, Proteus mirabilis, Morganella morgani, and Enterococcus faecalis. It was initially administered by three intramuscular (intragluteal) injections at weekly intervals. The first results of clinical trials with SolcoUrovac, which were performed in Europe, showed that the vaccine was protective against recurrent UTI (326). Current information about SolcoUrovac is not available as Solco Basel appears to be defunct and SolcoUrovac is unavailable in Europe at this time. However, another intramuscular polyvalent inactivated UTI vaccine, called StroVac (Strathmann AG, Hamburg, Germany), is apparently approved for use in Europe. StroVac contains the same 10 strains in a different formulation [http://www.strathmann.de/index.php/en/component/content/article/112-pflichtangaben/367-strovac-pflichttext-, reference in German]. While these vaccines have shown promise (261, 327), to our knowledge they have never undergone large phase III studies to demonstrate efficacy. However, the European Association of Urology’s “Guidelines on Urological Infections” mention SolcoUrovac and StroVac as options to consider in the non-antibiotic prophylaxis of recurrent UTI (http://uroweb.org/wp-content/uploads/18-Urological-Infections_LR.pdf).

Vaginal mucosal delivery of SolcoUrovac

In the initial human studies with intramuscular injections of SolcoUrovac, some women experienced adverse effects such as pain (5.4%), fever (3.5%) and swelling at the injection site (1.5%) (326). Thus, David Uehling, a pioneer in the field of UTI vaccines from the University of Wisconsin, tested the efficacy of vaginally administered SolcoUrovac, hypothesizing that mucosal administration in the vagina would reduce adverse effects. Vaginal instillation was effective in mice (328), cynomolgus monkeys (329), and women (330), paving the way for phase II clinical trials of vaginally instilled SolcoUrovac in the United States. SolcoUrovac’s phase II clinical trials with the vaginal suppository form of the vaccine, which were published between 1996 and 2007, were only partially successful (331–334). The most effective treatment course was determined to be six vaginal suppositories given at weeks 0, 1, 2, 6, 10, and 14. With this vaccination regimen, the percentage of women having a recurrence during the 6-month trial declined from 83% to 89% in the placebo-treated groups to 45% to 54% in the vaccinated, boosted groups. However, these differences were not always significant. In one trial, the authors identified six patient sub-populations with significantly lower re-infection rates after vaccination: “women younger than 52 years, without a childhood history of recurrent UTIs, [having] 6 or more UTIs in the previous year, without a hysterectomy, using estrogen, [or] using birth-control pills (331).” Adverse events (e.g., low-grade fever, nausea, vaginal irritation) did occur, but no patient was unable to complete the treatment.

The adaptive response to SolcoUrovac

After intramuscular vaccination, mice had 10-fold more total IgG and 2-fold more total IgA in the urine; IgM was not found in the urine (335). Vaginal immunization of monkeys was protective but did not increase anti-E. coli serum, vaginal wash, or urine antibody levels (serum IgG, IgM, and IgA; vaginal wash and urine IgA and IgG) (329). Interestingly, although some vaccinated women did have increased antibody titers over time, there were no significant differences in any of the tested antibody levels among treatment groups in any phase II trial (331–334). As expected by the formulation, SolcoUrovac was most effective against UPEC uropathogens. In one study, the percentage of women who experienced a UPEC UTI after vaccination and boosting was 27.5%, compared to 70% of women in the placebo group. However, the percentage of infections caused by uropathogens other than UPEC increased dramatically in vaccinated women and overall the vaccine did not statistically significantly prevent recurrent UTI when all uropathogens, not just UPEC, were considered (331). Perhaps because of this shift in uropathogens, to the best of our knowledge vaginal SolcoUrovac has not progressed to Phase III trials or beyond.

CP923

In 2007, Johnson and colleagues described a candidate vaccine consisting of a formalin-killed derivative of the E. coli sepsis strain CP9 that is deficient in capsule and
O-antigen, termed CP923 (336). Compared to formalin-killed CP9, intranasal vaccination with formalin-killed CP923 resulted in a significantly greater systemic-antibody response that was able to bind to a subset of heterologous UPEC and bacteremia strains. The mucosal immune response and protection against urinary-tract infection were not assessed, but this study shows the potential benefit of using genetically modified UPEC for inactivated vaccines.

Attenuated vaccines

Attenuated vaccines have the potential benefit of progressing through early steps in disease pathogenesis in the relevant host niche. Recently, the Klumpp group identified as a vaccine candidate a mutant of the UPEC strain NU14 that lacks the O-antigen ligase waaL (287). This gene was identified in a screen of transposon mutants that had lost the ability to suppress IL-8 production by bladder epithelial cells (337). In a murine-UTI model, NU14 ΔwaaL was significantly more inflammatory and less virulent than wild-type NU14. Vaccination with NU14 ΔwaaL protected mice from challenge infection with NU14, CFT073, and four UPEC isolates from the E. coli Reference Collection (287, 338). However, protection waned over time and was absent by 8 weeks. Interestingly, vaccination also significantly reduced the level of persistent bladder colonization (indicative of a QIR population) by NU14 14 days after challenge, even though the NU14 ΔwaaL (vaccine) strain itself is unable to persist past acute infection. The authors hypothesized that the lack of O antigen on LPS in NU14 ΔwaaL may allow for Toll-like receptor 4 recognition of LPS lipid A, or may increase the exposure of bacterial-surface antigens to antigen-presenting cells, thereby stimulating protective immunity (287). However, the recent finding that O antigen modulates infection-induced bladder pain, and that serial infections with NU14 ΔwaaL result in chronic bladder pain, diminishes the promise of NU14 ΔwaaL as a vaccine candidate (339).

Specific-Antigen Vaccines in Development

Specific-antigen vaccines (such as toxoid, conjugate, and subunit vaccines) have become more popular in recent years due to advances in protein purification and the development of recombinant-DNA technology. Single-antigen vaccines typically have lower rates of adverse events than whole-cell vaccines, and several antigens may be combined in a single multi-epitope vaccine to increase efficacy, e.g., in a recent extra-intestinal pathogenic E. coli vaccine (340). Candidate antigens may be revealed by UTI-virulence studies in vitro and in animal models; alternatively, reverse vaccinology allows researchers to predict effective antigens computationally.

Conjugate vaccines

Conjugate UTI vaccines against UPEC capsule and LPS components have shown protection in animal models after same-strain challenge, but have not been tested clinically in humans. In early studies, intraperitoneal injection of E. coli endotoxin protected rats from pyelonephritis (284), and the protection was later determined to be mediated by antibodies against O antigen (341). Subcutaneous and bladder injection of purified O antigen from E. coli O6 serotype protected rats from bladder infection with the same O6 strain (342). Decades later, O polysaccharide prepared from E. coli O8 LPS, conjugated to bovine serum albumin, reduced renal scarring and intratubular-neutrophil infiltration in rhesus monkeys that were challenged with an O8 UPEC strain (343). Purified E. coli K13 polysaccharide conjugated to bovine serum albumin protected rats from pyelonephritis (344). A different group conjugated E. coli K13 to diphtheria toxoid and found that the vaccine decreased renal bacteria load and disease-severity scores in mice after challenge with a K13 UPEC strain (345). A considerable challenge in formulating a vaccine targeting capsule or O antigen, the most exposed component of LPS, is the fact that there is a great heterogeneity of serotypes among E. coli isolates. For example, 6 different O serotypes account for only 75% of UPEC isolates (346), making the formulation of a broadly protective conjugate vaccine impractical. Furthermore, some capsule serotypes, such as K1, are thought to evade the host-immune response by molecular mimicry, potentially making them poor vaccine candidates (347).

Toxoid vaccines and outer-membrane vesicles

UPEC toxins have not been demonstrated to play a required role in UTI pathogenesis, so they are not ideal vaccine candidates. For example, a purified α-hemolysin toxoid vaccine prevented renal injury, but not colonization, in mice after challenge with a hemolytic UPEC strain (193). Rather than being secreted as “naked” proteins, UPEC toxins such as α-hemolysin and CNF1 are associated with outer-membrane vesicles (OMVs), which bleb from the surface of Gram-negative bacteria during all stages of growth (348). OMVs also contain adhesins, enzymes, and nonprotein antigens like LPS (348), and recently, a high-throughput tandem mass spectrometry approach was used to define the UPEC outer membrane proteome (349). OMVs may be
a mechanism for UPEC and other bacteria to protect their toxins while they are en route to host cells, and to deliver “concentrated bursts of effector molecules” to modulate host-cell processes (350). OMVs are intriguing vaccine candidates, and because they contain LPS and other pro-inflammatory virulence factors, they should not require adjuvants to stimulate the immune system. Several successful meningococcal-OMV vaccines have been developed, and other OMV vaccines have been effective in mice (351–355). UPEC OMVs are thus candidate vaccine antigens, though to our knowledge they have not been tested.

**Pili as vaccine candidates**

Pili are adhesive surface organelles that mediate the colonization of mucosal surfaces. Adhesins make an attractive antigen candidate, because antibodies raised against an adhesin should be able to block adhesin-host cell-receptor binding, thus disrupting bacterial colonization of the host (356). Several types of pili have been investigated as UTI vaccine candidates. Pilus vaccines have been tested since well before the effects of growth conditions on pilus production were fully understood, and as such, many early pilus vaccine papers do not describe the bacterial growth conditions for the challenge inocula. When possible, we will report the relevant information. To our knowledge, the first pilus-UTI vaccine was published in 1979. Rats that were vaccinated intradermally with pili purified from two clinical isolates of *E. coli* were protected from pyelonephritis; anti-pili antibodies raised in rabbits were also protective in rats (357).

**FimH**

Vaccines targeting the type 1 pilus adhesin FimH, which plays a critical role in UTI pathogenesis in the lower urinary tract in animal models, have been tremendously effective in animals that are challenged with bacteria grown in type 1 pilus-inducing conditions. Since the tip adhesin is functionally critical, but not highly abundant, purified adhesin was found to be better than whole pili at eliciting antibodies that blocked receptor binding (39). FimH can be purified bound to its periplasmic chaperone FimC, or as a naturally occurring, mannose-binding FimH truncate (FimHt) (358). Both antigens protected subcutaneously vaccinated mice from cystitis (39). The FimCH vaccine also protected intramuscularly injected cynomolgus monkeys from bacteriuria and pyuria though the number of animals tested was by necessity small (38). Of note, only one out of four FimCH-vaccinated monkeys developed bacteriuria and pyuria upon challenge infection with type 1 pilus-expressing UPEC (compared to four out of four control monkeys), and this was also the only FimCH-vaccinated monkey without increased anti-FimH IgG in vaginal secretions. Another group compared the efficacy of a recombinant FimHt vaccine administered either intranasally or intramuscularly to mice. Both routes were protective against cystitis, but the intranasal vaccine induced greater anti-rFimHt IgA in vaginal washes (300). Yet another group demonstrated that subcutaneous administration of recombinant fimH fused to the flagellin subunit fliC, a TLR5 agonist and candidate adjuvant, protected mice against cystitis upon challenge with a type 1 pilus-expressing clinical isolate; vaccination with admixed FimH, FliC, and Montanide ISA 206 adjuvant was also protective (359, 360). These investigations of cellular immune response to vaccination are unique among UTI vaccine studies, which generally test the humoral response only. Immunization resulted in Th1 and Th2 responses as assessed by cytokine responses in splenocyte-proliferation assays and ratio of IgG1 to IgG2a. Lastly, another group has recently developed a mammalian codon-optimized fimH plasmid construct for use in a DNA vaccine, whereby plasmid DNA injected into mice can induce a protective immune response (361, 362). Mice that received the DNA vaccine via footpad injection had significantly reduced bladder colonization 48 hours post-challenge and significantly increased urine IgA titers (363).

While it has been reported that monoclonal antibodies raised against FimH do not block, but rather enhance, adherence to bladder epithelial cells in vitro (364), the above studies demonstrate that polyclonal IgG raised against FimH block bacterial binding to bladder epithelium and is clearly protective in vivo. Possible explanations for this discrepancy include steric hindrance preventing the antibody-adhesin complex from binding to the uroplakin-receptor pocket and the effects of opsonization. The FimCH vaccine was originally licensed by MedImmune (Gaithersburg, Maryland, USA) and entered Phase I and II trials in the early 2000s. The vaccine was found to be safe in Phase I trials, but was dropped from development during Phase II trials. Sequoia Sciences (St. Louis, Missouri, USA) has since acquired the license and the vaccine is re-entering clinical trials in women with recurrent UTI, using a new adjuvant. In two pre-clinical rabbit studies conducted by Sequoia, serum IgG anti-FimH titers were greater than 1:1,000,000, with no apparent adverse effects from the vaccination (personal communication, Gary Eldridge, Sequoia Sciences).
P pilis
P pilus-subunit vaccines to protect against pyelonephritis became a hot topic in the 1980s and the initial studies showed promise. Vaccination with purified recombinant P pilis blocked renal colonization in mice when the challenge bacteria were grown under P pilus-inducing conditions (37, 365). Vaccination with purified P pilis protected monkeys (366, 367) and the unvaccinated infants of vaccinated monkeys (298) from pyelonephritis when the challenge bacteria were grown under P pilus-inducing conditions. Finally, synthetic P pilus peptides that were prepared by solid-phase Merrifield synthesis and conjugated to carrier proteins prevented urine and renal colonization in mice (368). However, studies utilizing whole-purified P pilus-UTI vaccines have not been published since the late 1980s. This is likely due to the high degree of antigenic variation among UPEC strains in the major P pilin subunit, PapA, which is the most abundant pilin protein in P pilus preparations. Indeed, natural P pilus-specific antibodies from patients with pyelonephritis do not seem to target the binding pocket of PapG as they are unable to prevent P pili-mediated hemagglutination (96). Consistent with this, an inactivated whole-cell vaccine consisting of formalin-killed P-fimbriated E. coli offered only limited protection against renal dysfunction and scarring in monkeys (325). Thus, whole-cell vaccines may not be an effective way of inducing anti-pilin antibodies, even if they are being expressed on the bacterial surface.

PapDG vaccine
By 1988, the composition of P pilis had been determined, and the tip-adhesin protein PapG was identified as a vaccine candidate (61, 369). Lund and colleagues suggested that PapG could be purified in a complex with its periplasmic-chaperone protein PapD, analogous to the FimCH vaccine. In 2004 it was shown that intraperitoneal administration of a purified PapDG vaccine protected cynomolgus monkeys from pyelonephritis (40). The efficacy was presumed to be the result of PapG-specific antibodies blocking the pilus adhesins and thereby preventing colonization, though the specific mechanism of protection is unknown. To the best of our knowledge, no further studies have been conducted with the PapDG-pyelonephritis vaccine.

Other pili
Among the adhesins expressed by some UPEC strains are S pilis and Dr adhesins (and others), each of which has been used as a vaccine antigen. Rats vaccinated with purified recombinant S pilis had reduced kidney colonization (370). In the same study, an avirulent strain of Salmonella enterica serovar Typhimurium was genetically transformed to produce S pilis, and live bacteria were orally administered to rats, which had reduced kidney colonization compared to mock-vaccinated and purified S pilus-vaccinated mice (370). In addition, mice vaccinated with purified recombinant Dr adhesins had reduced UTI-associated mortality (371). However, these adhesins are even less broadly conserved among UPEC than are P pili antigens, and thus, these targets would only be useful in a multi-epitope vaccine. Pilus antigens from non-UPEC uropathogens have shown efficacy in mouse models of infection, as described below.

Subunit vaccines and reverse vaccinology
Recently, investigators have used information gathered through bioinformatic, genomic, and proteomic analyses to identify novel candidate antigens, in an approach termed “reverse vaccinology.” The first web-based reverse-vaccinology program, Vaxign, was used to predict 22 UPEC outer-membrane proteins as potential vaccine targets (372). Some of these targets had been previously shown to be immunogenic and protective in animal models, while others remain to be tested. A large-scale reverse-vaccinology screen was used to identify vaccine-antigen candidates in E. coli CFT073, which is predicted to encode 5379 proteins. The criteria for candidate antigens were pathogen specificity, high in vivo expression, induction during growth in human urine, antigenicity, and surface exposure. Six candidates were identified, each an outer membrane-receptor protein involved in bacterial iron or heme acquisition. When purified and administered intranasally, the candidate antigens Hma, IreA, and IutA protected mice from challenge infection (373). Vaccination with Hma, a heme receptor, protected the kidneys; IreA, a putative siderophore, protected the bladder; and IutA, a siderophore receptor for aerobactin, protected both the bladder and the kidneys. The three antigens also induced antigen-specific IgA in the urine and class-switching from IgM to IgG in the serum (373). A subsequent study investigated additional UPEC outer membrane iron receptors as vaccine candidates. The yersiniabactin receptor FyuA, purified and administered intranasally, protected mice from developing pyelonephritis upon challenge with 536, a UPEC strain that expresses FyuA. Vaccination-elicited anti-FyuA IgA in urine and IgG in serum, and serum-antibody levels were correlated with kidney bacterial burden (374). A recent RNAseq analysis confirmed that the yersiniabactin system is highly expressed by UPEC during uncomplicated cystitis in women (375).
Another group had previously found that systemic vaccination with the siderophore-receptor IroN protected against renal colonization in mice (180). This last study did not explicitly use a reverse-vaccinology approach, but IroN was chosen because of its prevalence among clinical UPEC isolates, its role in urovirulence, and its expression in bodily fluids.

Another reverse-vaccinology approach employed by a group at Novartis (Siena, Italy) involved comparing the genome of a neonatal meningitis-associated K1 strain of E. coli with known pathogenic and nonpathogenic E. coli strains. Potential antigens were chosen if they were predicted to be surface-associated or secreted, with no more than three transmembrane domains, and were absent from nonpathogenic strains. Two hundred and thirty candidates were identified in this manner and tested for protection in a murine-sepsis model; nine were protective (376). One protective antigen, named FdeC for factor-adherence E. coli, was found to be expressed by most UPEC, but only upon host cell contact, helping to mediate E. coli adhesion to mammalian cells. Intranasal vaccination with recombinant FdeC significantly reduced kidney colonization in mice that were challenged with UPEC strains 536 or CFT073 (122). Another protective antigen, SseE (secreted and surface-associated lipoprotein from E. coli; also known as YghJ), was found to be involved in the degradation of mucin substrates (377). Intranasal vaccination with recombinant SseE significantly reduced kidney and spleen colonization in mice that were challenged intravesically with the UPEC strain 536, which expresses a different variant of SseE.

**Vaccines Against Non-UPEC UTI**

Uropathogenic E. coli cause approximately 85% of uncomplicated UTI, and so it is not surprising that most tested UTI vaccines have used UPEC strains and antigens. However, vaccines targeting other uropathogens have been protective in animal models. The polyvalent inactivated vaccine SolcoUrovac/StroVac (described in detail above) contains one strain each of Klebsiella pneumoniae, Proteus mirabilis, Morganella morganii, and Enterococcus faecalis. Vaginally instilled, formalin-killed K. pneumoniae inhibited bacterial adherence to rat bladder mucosae (323). In addition, a recent immunoproteome analysis identified candidate antigens for a K. pneumoniae vaccine (378), but to the best of our knowledge, these antigens have not been tested in a UTI model. Most of the other non-UPEC vaccines have targeted P. mirabilis, which causes about 3% of uncomplicated and 13% of complicated UTI (379).

**Proteus vaccines**

Vaccines against P. mirabilis infection have been tested since at least the 1960s, when heat-killed P. mirabilis protected rats from pyelonephritis by promoting renal clearance of bacteria (380). A preparation of P. mirabilis outer-membrane protein promoted renal clearance in mice and protected mice from renal infection and death (379). Purified inactivated-Proteus toxic agglutinin (Pta), a cytotoxic surface-associated alkaline protease, was conjugated with cholera toxin and protected mice from kidney colonization (381). Finally, several P. mirabilis adhesins have been tested as vaccine antigens. P. mirabilis expresses MR/P (mannose-resistant, Proteus-like) fimbriae on the cell surface, and most of the bacterial population synthesizes MR/P fimbriae during UTI (382). Vaccination with purified MR/P fimbriae was protective against ascending P. mirabilis UTI in a murine model (383). An attenuated-mucosal vaccine consisting of Lactococcus lactis expressing the recombinant MrpA subunit of MR/P fimbriae significantly reduced renal colonization in mice after P. mirabilis challenge (384). Systemically injected, purified recombinant MrpA also protected mice from ascending P. mirabilis UTI (385). Intranasal vaccination with recombinant MrpA protected mice from ascending P. mirabilis UTI (386, 387); the addition of a cholera toxin adjuvant did not enhance protection (388). MrpH is the MR/P fimbrial-tip adhesin, similar to FimH and PapG (described above). Vaccination with recombinant MrpH was protective against ascending P. mirabilis UTI in a murine model (383). A fusion protein comprised of recombinant MrpH and UPEC FimH protected intranasally-vaccinated mice from challenge with either P. mirabilis or UPEC (389). Other P. mirabilis antigens have also been tested. Vaccination with the urothelial cell adhesin subunit UcaA protected mice from P. mirabilis infection in a hematogenous infection model (385). However, intranasal vaccination with flagellin was not protective in mice. Interestingly, flagellin co-administered with MrpA negated the protective effect of MrpA vaccination, suggesting an immunomodulatory effect (387).

**Enterococcus vaccines**

The endocarditis- and biofilm-associated pilus (Ebp pilus) (Fig. 2) is an attractive vaccine candidate. Recently, the minor pilus subunit EbpA was found to be an adhesin that binds to fibrinogen, a host protein that is released upon bladder catheterization (216). Systemic vaccination of C57BL/6 mice with E. faecalis EbpA prior to catheter implantation and E. faecalis infection significantly reduced bacterial colonization of the catheter.
and protected against CAUTI. Furthermore, serum from vaccinated mice was found to block EbpA binding to fibrinogen in vitro.

Immunotherapeutic Compounds
OM-89/UroVaxom

The immunotherapeutic formulation OM-89 (marketed in Europe by EurimPharm GmbH as UroVaxom) is a bacterial extract prepared from 18 strains of E. coli. For the purpose of preventing recurrent UTI, it is administered orally, typically as a daily dose for three months, and is recommended by the European Association of Urology for women with recurrent uncomplicated UTI. Several meta-analyses have assessed the effectiveness of OM-89 in preventing recurrent UTI in humans (261). A meta-analysis of five placebo-controlled double-blind studies found that OM-89 was superior to placebo with regards to reducing UTI frequency and dysuria, bacteriuria, and leukocyturia (390). Another meta-analysis of five studies found that the mean number of UTI episodes and the use of antibiotics were reduced in patients treated with OM-89 (391). OM-89 is generally safe and well-tolerated; the most frequent adverse events are headache and gastrointestinal events. While early studies looked at UTI recurrence over just six months from the start of treatment, OM-89 is effective for up to 12 months when booster doses are administered for the first 10 days in months 7, 8, and 9 (392). Several other immunostimulatory compounds exist for the prevention of recurrent UTI. Uromune, a preparation of E. coli, K. pneumoniae, P. vulgaris, and E. faecalis, was recently evaluated in a multicenter, retrospective, observational study. Women with a history of recurrent UTI who received daily Uromune prophylaxis for three months had significantly fewer recurrences over a 15-month period than women who received daily trimethoprim-sulfamethoxazole for six months (393). Other similar formulations, such as Urostim and Urvakol, have been developed, but few controlled studies are available and they will not be discussed further here.

The mechanism by which immunostimulatory compounds protect against recurrent UTI remains unclear. In mice, OM-89 activated macrophages (394) and induced a T-helper-1-type immune response as determined by increased IgG2a in the serum and interferon (IFN)-γ in spleen-cell supernatant (395). OM-89 also increased interleukin (IL)-6 and IFN-γ levels and decreased inflammation in the mouse bladder (396). The antibody response to OM-89 varies in different studies. In mice, OM-89 mainly induced IgG in the serum, and increased IgM only weakly (397). Also in mice, strain-specific IgG and IgA were increased in immune sera, and total and strain-specific IgG and IgA were increased in the urogenital tract. In addition, the antisera could recognize other human uropathogens such as E. faecalis, K. pneumoniae, and P. mirabilis. However, cross-reactivity was stronger with intraperitoneal injection of OM-89, rather than oral administration (395, 398). Interestingly, in a meta-analysis of two studies, there was no significant difference in urine and vaginal fluid anti-E. coli IgG and IgA between OM-89-treated and placebo-treated patients (391), which would suggest that OM-89’s efficacy may not be antibody-mediated. As OM-89 is a lysate of E. coli administered daily for several months, its efficacy could be the result of induced LPS tolerance. Similarly, a TLR4 polymorphism that is associated with reduced inflammatory signaling in response to LPS is also associated with protection from recurrent UTI (40).

CONCLUSIONS

The Challenge is Great

The progress in our understanding of UTI pathogenesis over the past 15 years has been truly remarkable and has begun to change how UTI is viewed and treated in the clinic. Sophisticated animal models of infection and translational studies have revealed that, rather than being a simple extracellular infection of the urinary tract mucosa, infection with UPEC and a number of other Gram-negative uropathogens (which together cause more than 80% of all UTI) proceeds through dynamic intracellular and extracellular-host niches during the course of acute and chronic infection. Whole-genome sequencing and gene-expression analysis of uropathogens have allowed an unprecedented look into the lifestyle of these versatile pathogens, thus enabling genetic and computational approaches to identify novel virulence mechanisms and vaccine candidates. The determination of structural details of uropathogenic adhesins has led to the rational design of anti-infectives and preventative therapies. However, with increased understanding comes knowledge of the imposing challenges facing scientists in the effort to develop broadly protective therapies. These challenges stem from the fact that the development of symptomatic UTI is exceedingly complex, hinging upon two factors that are highly variable and yet define the host-pathogen interaction: the virulence of the infecting uropathogen and the character of the bladder mucosal-immune response. On the one hand, uropathogens are a very heterogeneous collection of bacterial isolates that have the capacity to inhabit...
diverse host niches, including the gut, the urogenital tract, the bloodstream, and the meninges. Therefore, even among isolates from a single species (such as UPEC), uropathogens differ widely in their genetic and epigenetic makeup, and thus are represented by a large and varied number of serotypes and virulence-factor profiles. On the other hand, the host genetic and environmental variables that determine the extent and character of the bladder mucosal-immune response to infection are considerable and very poorly understood. As a result, two individuals may be infected with the same strain but have very different responses to infection, ranging from an asymptomatic-carrier state and/or asymptomatic bacteriuria, to severe cystitis and pyelonephritis with renal scarring (35).

**A Call to Arms: Investigators in Bladder Mucosal Immunity Needed!**

Developing new and efficacious therapies should be the highest priority in UTI research, as such therapies have the real potential to positively affect the quality of life of millions of individuals and decrease the overall use of antibiotics. However, many challenges must be overcome. The significant contribution of the host to recurrent UTI makes it unlikely that any new vaccine or therapeutic alone will completely eliminate recurrent UTI in all patients, unless it is also able to alter the innate mucosal-immune response to uropathogens. As the clinical trials of the vaginal vaccine Urovac have demonstrated, even a whole-cell vaccine broadly targeting several UPEC serotypes, as well as four other uropathogens, was only able to significantly protect against recurrent UPEC UTI, and then only in a subset of women. Importantly, what the researchers saw in many of these women was a shift towards recurrent UTI caused by less common uropathogens, suggesting a defect or sensitization of the mucosal-immune response. In the absence of vaccination, UPEC were able to predominate in the urogenital niche of these subjects, but with vaccination, other uropathogens replaced UPEC (331). Thus, as new and more efficacious strategies are being developed to combat recurrent UTI by UPEC, such as pilus-adhesin antagonists (e.g., mannosides) and UPEC-subunit vaccines, we must anticipate the likelihood that a subset of patients will continue to suffer from recurrent UTI with less-common uropathogens. For these patients, therapeutic interventions targeting the bladder mucosal immune response may provide additional benefit and relief from symptoms. However, our understanding of bladder-mucosal immunity is currently insufficient to allow informed predictions, and there is a paucity of investigators in this field. Despite the pioneering efforts of Dr. Svanborg and others, the field is relatively small. For example, of the 10 investigator teams of the Mucosal Immunity Study Team National Institutes of Health (NIH)-National Institute of Allergy and Infectious Diseases (NIAID) U01 consortium (www.mucosal.org), only one group is focused on investigating the urogenital tract. Until our understanding of bladder mucosal immunity matures, which will require a critical mass of investigators in this field, novel approaches to the treatment and prevention of UTI may be slow in coming.

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Conflicts of interest: Scott Hultgren is a part owner of Fimbrion and may financially benefit if the company is successful in marketing the mannosides, pilicides, and curlicides that are related to this article. He may also receive royalty income based on the FimH vaccine technology that he developed, which was licensed by Washington University to Sequoia Sciences.

**REFERENCES**

Drug and Vaccine Development for UTI


Drug and Vaccine Development for UTI


Immun 54: nephropathogenicity in an experimental rat pyelonephritis model.

virulence factor in murine urinary tract infection.

Escherichia coli pathogenic induced antigens including an RTX family exoprotein required for uro-

One 8:e61169.

HL, Tarr PI. urinary tract infections.


117. Spurbeck RR, Stapleton AE, Johnson JR, Walk ST, Hooton TM, Mobley HL. 2011. Fimbrial pro-


O’Brien et al.


215. Flores-Mireles AL, Pinisper NS, Caparon MG, Hultgren SJ. 2014. EbpA vaccine antibodies block binding of Enterococcus faecalis...
to fibrinogen to prevent catheter-associated bladder infection in mice. Sci Transl Med 6:254ra127.


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